

Benthic processes in fresh water fluffy sediments undergoing resuspension

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ABSTRACT

In the Po river plain relict freshwater wetlands are characterised by a low free water to emergent macrophyte surface ratio, rapid infilling and fluffy sediments, undergoing frequent resuspension. Particle mixing should alter the steep gradients of dissolved gas, nutrient and organic matter quality that generally characterise sediments, with implications for benthic processes.

Sediment features and solute fluxes were studied from December 2003 to February 2005 within the Busatello swamp complex (Northern Italy) by means of a combination of core incubation, porewater extraction and microprofiling. At the study site, along a 10 cm vertical profile, sediment organic matter content (32.5%), porosity (0.94) and density (1.02 g cm^{-3}) were nearly constant.

Oxygen demand measured by dark core incubation ($12.7\text{-}56.9 \text{ mmol m}^{-2} \text{ d}^{-1}$) was strongly correlated with water temperature. Rates agreed reasonably well with diffusive oxygen fluxes calculated from microprofiles while ammonium and phosphorus regeneration rates predicted from porewater gradients ($-309.3\text{-}43.4$ and $-0.1\text{-}0.7 \text{ } \mu\text{mol m}^{-2} \text{ d}^{-1}$, respectively) were significantly lower than rates measured via core incubations ($-129.8\text{-}5420.5$ and $-120.4\text{-}35.4 \text{ } \mu\text{mol m}^{-2} \text{ d}^{-1}$, respectively). This is a probable consequence of insufficient vertical resolution of nutrient profiles and lack of steady state conditions. A sediment resuspension experiment, carried out under controlled laboratory conditions, indicated large instantaneous oxygen and nitrate consumption, a transient increase of denitrification rates and the rapid release of ammonium and soluble reactive phosphorus. In unconsolidated sediments, resuspension and mixing of sediment particles are key processes regulating mineralisation rates and benthic-pelagic coupling.

Key words: unconsolidated sediments, resuspension, porewater profiles, core incubations, oxygen demand, denitrification.

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INTRODUCTION

In shallow aquatic environments, processes at the sediment-water interface have a well recognised role as regulators of many solute dynamics (Emerson, 1976; den Heyer and Kalfs, 1998). Benthic aerobic respiration and chemical oxidation modulate bottom water oxygen availability and related processes (*i.e.*, nitrification rates) and can result in the exhaustion of this electron acceptor (Froelich *et al.*, 1979; D'Angelo and Reddy, 1994a, 1994b). Ammonium and phosphorus recycling in sediments and subsequent release to the overlying water has the potential to fuel water column primary production and maintain long-term eutrophication conditions (Blackburn and Henriksen, 1983; Mayer *et al.*, 1999; Soto-Jimenez *et al.*, 2003).

A considerable number of benthic flux measurements have been performed in a wide variety of nearshore marine environments (Callender and Hammond, 1982; Hopkinson and Wetzel, 1982; Archer and Devol, 1992; Rasmussen and Jorgensen, 1992; Rysgaard *et al.*, 1995; Rabouille *et al.*, 2003), in the open ocean (Glud *et al.*, 1998; Jørgensen *et al.*, 2005) and in lakes (Sweerts *et al.*, 1991a; den Heyer and Kalfs, 1998; Liikanen *et al.*, 2003), while, to our knowledge, only a few studies were carried out in fluffy sediments of freshwater wetlands (Jensen and Andersen,

1992; Reddy *et al.*, 1996; Mayer *et al.*, 1999; Fisher and Reddy, 2001; Longhi *et al.*, 2008). Here, oxygen dynamics and nutrient regeneration rates are strictly coupled and affect a variety of wetland functions and processes, such as primary production, greenhouse gas emission and the sedimentary accumulation of organic matter (Longhi *et al.*, 2008; Pierobon *et al.*, 2010). Uncoupling between high sedimentation of organic matter and mineralisation rates determines the burial of refractory material, the accumulation of fluffy sediments and ultimately the shift from shallow aquatic environments towards terrestrial ecosystems. Detailed knowledge of decomposition rates and nutrient recycling in these systems can thus provide helpful information for management purposes (Godshalk and Wetzel, 1978; Reddy and D'Angelo, 1994).

In consolidated sediments, benthic solute exchange is regulated by abiotic and biotic factors such as concentration gradients, sediment tortuosity, microbial and benthic microalgal activities, burrowing, ventilation or excretion activities by macrofauna (Aller, 1980; Berner, 1980; Blackburn and Henriksen, 1983; van Der Loeff *et al.*, 1984; Kristensen, 1985; Sundback *et al.*, 1991; Sweerts *et al.*, 1991a; Rizzo *et al.*, 1992; Rysgaard *et al.*, 1995). Instead, in unconsolidated sediments, solute transfer across the sediment water interface may be strongly influenced also by

resuspension of sediment particles induced by wind, bioturbation by large fishes and methane ebullition (Breukelaar *et al.*, 1994; Liikanen *et al.*, 2003; Haeckel *et al.*, 2007).

Different methods were used to determine the magnitude and direction of solute fluxes across the sediment water interface: *in situ* incubation of benthic chambers (Rowe *et al.*, 1975; Callender and Hammond, 1982; Hopkinson and Wetzel, 1982; van Der Loeff *et al.*, 1984), laboratory incubation of intact sediment cores (Rasmussen and Jorgensen, 1992; Moore *et al.*, 1998) and extraction or microsensor profiling of porewaters coupled to deterministic models of solute flux, such as Fick's laws of diffusion (Berner, 1980; Klump and Martens, 1981; Sweerts *et al.*, 1991a; Berg *et al.*, 1998). The results from these approaches to flux measurement are often compared in order to obtain some estimate of the variability between the techniques, and in the attempt to determine the most appropriate experimental approach for a given system.

In this study we evaluated dark oxygen demand and ammonium and phosphorus regeneration rates over a one-year period by means of intact core incubation, microprofiling and porewater extraction in unconsolidated wetland sediments. We also simulated sediment resuspension and evaluated its effect on oxygen demand, denitrification rates and nutrient fluxes. The investigation had a threefold aim: i) analyse the seasonal evolution of benthic mineralization rates in relation to water temperature, ii) analyse the effect of sediment resuspension and iii) compare re-

sults obtained with different commonly used techniques for quantifying benthic processes.

We hypothesise that in unconsolidated sediments transient episodes of resuspension and particle mixing i) enhance nutrient regeneration and promote the mineralisation of organic matter and ii) set frequently to zero solute gradients and impede steady state conditions of the benthic system.

METHODS

Study area

This study was conducted from December 2003 to February 2005 in the Natural Reserve *Paludi del Busatello*, an 81 ha wetland complex located in between the provinces of Mantua and Verona, Northern Italy (Fig. 1). This marsh is a residual area of a vast wetland system that was almost entirely converted into agricultural land in the last decades.

Most of the surface is colonized by reed stands (67 ha); the former Busatello river (which is now a narrow shallow channel with almost stagnant water) divides the Reserve in two areas having approximately the same surface; besides the Busatello river, the marsh hosts a network of ponds. The wetland environment is undergoing rapid burial due to accumulation of refractory organic matter and active management is required to protect the relevant aquatic species as the turtle *Emys orbicularis*, the

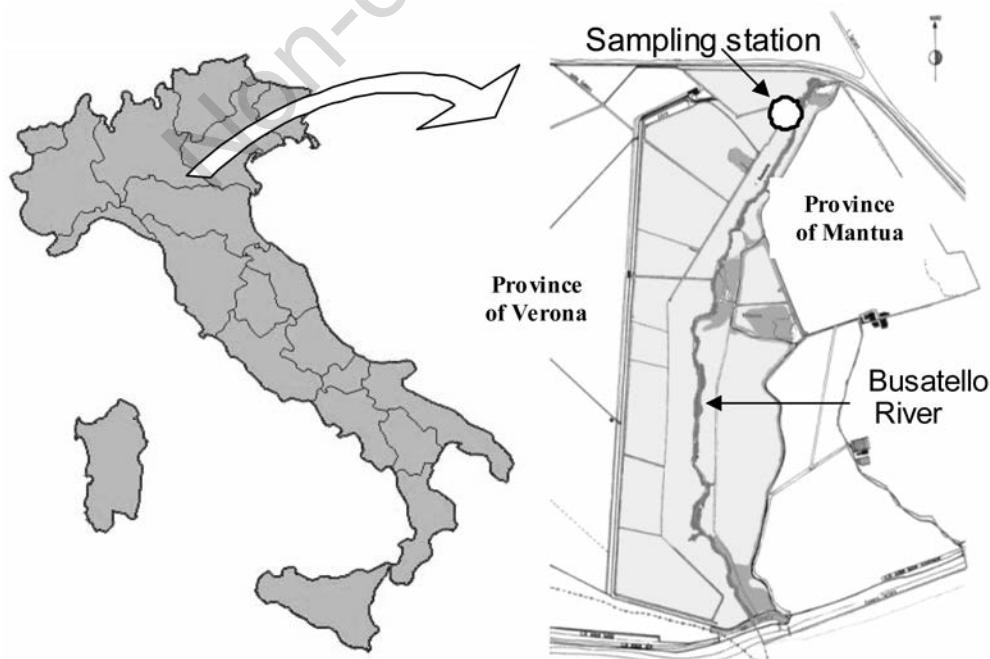


Fig. 1. Map of the Natural Reserve *Paludi del Busatello* with the indication of the sampling station.

endemic frog *Rana latastei*, a number of floating leaved and submersed macrophytes and, among the autochthonous fish, the pike *Esox lucius*.

Dominant macrophyte communities consist of the common reed *Phragmites australis*, sedges as *Carex riparia*, floating leaved plants as *Nuphar luteum* and *Nymphaea alba* and submersed plants as *Ceratophyllum demersum*, *Utricularia australis*, *Myriophyllum spicatum* and *Najas marina*. The site chosen for this study is located in the northern and more preserved part of the Reserve (Fig. 1), where the Busatello river enlarges and forms a pond which is devoid of macrophytic vegetation. Chlorophyll *a* (Chl *a*) generally peaks in winter ($\sim 20 \mu\text{g L}^{-1}$), while it is very low during summer months, resulting in transparent waters. Summer months are also generally characterised by low nutrient concentrations (dissolved inorganic nitrogen, $\text{DIN} < 25 \mu\text{M}$, soluble reactive phosphorous, $\text{SRP} < 0.1 \mu\text{M}$). Water depth is 1 m, sediments are fluffy and poor of macrofauna.

Sampling

Water and sediment samples were collected on 10th December 2003, 16th March, 21st July and 16th September 2004, and 23rd February 2005. On each date, water column temperature, pH, conductivity and dissolved oxygen concentrations were measured *in situ* with a multiple probe (ISI Instruments, model 556, Twin lakes, WI, USA) while DIN, SRP and Chl *a* concentrations were analysed in the laboratory with standard spectrophotometric methods (Koroleff, 1970; Valderrama, 1977; A.P.H.A., 1981). Six intact sediment cores were collected by hand using transparent Plexiglas[®] liners, and ~ 100 L of water were collected for core maintenance during transport, preincubation and incubation periods. Three cores (i.d. 20 cm, height 40 cm) were collected for flux measurements via dark laboratory incubation and 3 cores (i.d. 8 cm, height 40 cm) for sediment microprofiling and porewater characterisation by slicing and squeezing. In September 2004, 12 additional cores (i.d. 8 cm, height 40 cm) were collected for a sediment resuspension experiment; six cores were used for flux measurements and six for the determination of denitrification rates (see later).

After sampling, sediment level inside collected liners was adjusted to 20 cm leaving an overlying water column of about 10 cm. Cores were sealed in the bottom with rubber bungs and stored with the top open submersed in a box with cooled pond water. Within 4 hours from sampling, cores were transferred in a large tank containing aerated pond water at ambient temperature and left overnight. Water mixing inside cores was ensured with small pumps (20 cm i.d. cores) or Teflon[®]-coated magnetic bar suspended above the sediment interface and driven by an external motor (8 cm i.d. cores); water stirring was on during the whole preincubation and incubation periods.

Sediment characterisation and profiling

The day after the sampling, O₂, pH and redox potential (Eh) profiles were measured in triplicate in each core by means of micro and minisensors. O₂ and Eh profiles were done using microsensors with a 50 μm tip and a response time of ~ 0.3 and ~ 30 s, respectively (Unisense, Aarhus, Denmark). A saturated calomel electrode was used as reference sensor for Eh measurements (Radiometer, Brønshøj, Denmark). Profiles of pH were done using mini electrodes with tip 1 mm and response time of ~ 30 s (Diamond General, Ann Arbor, MI, USA). All sensors were mounted on a manual micromanipulator (vertical resolution 0.01 mm, Tesa Hite, Switzerland). Profiles had different vertical resolutions depending upon the parameter, the season and the diameter of the sensor. In particular, vertical resolution of oxygen profiles varied from 0.2 (July and September 2004) to 0.5 mm (December 2003, March 2004 and February 2005), while vertical resolution of Eh and pH profiles was 2 mm at each sampling date. The water-sediment interface was recognised as the layer with the steeper O₂ drop (Revsbech, 1983); O₂ profiles were limited to the oxic horizon, while pH and Eh profiles were conducted over the upper two sediment centimetres.

Immediately after profiling, the cores were extruded and sediments were sliced into 0-1, 1-2, 2-3, 3-5 and 5-10 cm intervals for density, porosity, water, organic matter, total C, N and P content determination and for porewater characterization (NH₄⁺ and SRP); interstitial water was obtained by squeezing the slices under N₂ atmosphere.

Sediment density was determined by weighing 5 cm³ of homogenised fresh sediment into pre-weighed ceramic dishes. The samples were then dried at 60°C to constant weight and reweighed to determine sediment porosity (mL H₂O mL sed⁻¹) and water percentage as weight loss. Organic matter content (loss on ignition, LOI) was evaluated from weight loss after ignition of dried and powdered sediment (~ 0.1 g) at 400°C for 3 hours. Dried sediments were also analysed for organic C and total N content (CHNS-O EA 1108, Carlo Erba Elemental Analyzer, Milano, Italy) and total P content according to Aspila *et al.* (1976).

Calculation of the diffusive fluxes of O₂, NH₄⁺ and SRP across the sediment-water interface was based on application of Fick's law to the concentration profiles as described by Berner (1980):

$$J = \frac{\delta C}{\delta z} \cdot D \quad (\text{eq. 1})$$

where J ($\mu\text{mol m}^{-2} \text{h}^{-1}$) is the flux, D ($\text{cm}^2 \text{s}^{-1}$) is the diffusion coefficient and $\delta C/\delta z$ ($\mu\text{M cm}^{-1}$) is the concentration gradient. Molecules and ions diffuse according to the parameter D only in the water column and in the diffusive

boundary layer; within the sediments, D is generally corrected for tortuosity using the porosity and porosity-tortuosity relationship (Sweerts *et al.*, 1991a):

$$D_s = \phi^m \cdot D \quad (\text{eq. 2})$$

where D_s is the diffusion within sediments, ϕ is the sediment porosity and m is an empirical factor depending on sediment features.

For O_2 flux calculations we used the average porosity values relative to the upper sediment layer, while for NH_4^+ and SRP fluxes we used the average porosity calculated along the 0-10 cm horizon; for m we assumed a value of 1 as was done by other authors working with very soft freshwater sediments (Urban *et al.*, 1997). Diffusion coefficients were taken from Broecker and Peng (1974) and Li and Gregory (1974) and corrected for water temperature according to Li and Gregory (1974).

Estimation of sedimentation rates

Gross sedimentation rates were evaluated in March and September 2004 by means of cylindrical sediment traps, consisting of Plexiglas® liners (i.d. 5 cm, height 30 cm) (Bloesch and Burns, 1980). For each date, six traps were randomly placed on the bottom of the sampling station and retrieved after 23 (March) and after 15 days (September). Longer deployment in March coincided with minimum water temperatures and sedimentation rates and was set in order to accumulate enough material for reliable estimate of rates. We acknowledge that long traps deployment can lead to slight underestimate of sedimentation rate, but in this specific case microbial activity was limited by low water temperatures and by the quality of the organic matter. September exposure time overlapped that suggested by Bloesch and Burns (1980).

In the laboratory the trap contents were dried at 60°C and weighed. The sedimentation rate ($g\ m^{-2}\ d^{-1}$) was calculated as the dry mass of sediment accumulated normalised by the area of the trap and divided by the number of days that the trap was deployed.

Sediment core incubations

The day after sampling, cores were incubated in the dark for the determination of O_2 , NH_4^+ and SRP net fluxes. The morning after the sampling, water inside incubation tank and inside cores was replaced with fresh water. The incubation started just after, when floating Plexiglas® lids were positioned on the top of each core. Employed lids were perfectly fitting the inner core wall, minimising any significant gas exchange with the atmosphere during the incubation. At the beginning of the incubation, water samples were collected with a plastic syringe from the tank and then, at different time intervals, from a valve on the

top lids. Each time ~100 mL were collected; samples for O_2 analyses were immediately transferred into 12 mL glass tubes (Exetainers, Labco, Lampeter, UK) and analysed by Winkler titration (A.P.H.A., 1981). Samples for nutrient analyses were filtered (pore size 0.7 μm , GF/F Whatman filters, Whatman International Ltd, Maidstone, Kent, UK) and transferred into glass tubes for SRP determination or frozen in plastic vials for NH_4^+ analysis, as previously described. Total incubation time was comprised between two (July) and five (February) hours; fluxes were determined according to the following equation:

$$Fx = \alpha \cdot h \cdot 10 \quad (\text{eq. 3})$$

where Fx ($\mu mol\ m^{-2}\ h^{-1}$) is the flux of the x species, α ($\mu M\ h^{-1}$) is the slope of the linear regression between concentration of the x species and incubation time, h (cm) is the height of the overlying water column and 10 is a factor necessary to convert units.

Sediment resuspension experiment

In September 2004, 12 cores were collected for two sediment resuspension experiments, one for the measurement of oxygen and nutrient fluxes (6 cores) and one for denitrification (6 cores).

Flux measurements

Three cores were left untreated (C), while the sediment of three cores (upper 10 cm) was gently resuspended in the water column by means of a trowel to simulate a resuspension event (Mix). In Mix cores, samples of water overlying sediment were collected before and after sediment mixing, when particles were settled, and analysed for dissolved oxygen, NH_4^+ , NO_3^- and SRP concentrations. After water sampling, the water overlying sediments in Mix cores was carefully replaced with *in situ* water and thereafter C and Mix cores were incubated for the determination of O_2 , NH_4^+ , NO_3^- and SRP fluxes, according to the procedure previously described. We replaced the water overlying sediments as resuspension releases pore water solutes that deeply alter the chemistry of the core water. This has implication for gradients that do not reflect what happens in the natural environment. Here, released solutes are diluted in a large water volume and concentrations do not change appreciably. Our intention was to measure processes driven by porewater and sediment features altered by a short term resuspension event and for this reason we replaced overlying water with *in situ* water.

Denitrification measurement

$^{15}NO_3^-$ was added to the water column of the second set of 6 cores to increase the nitrate pool by ~30% and then three cores were mixed as previously described (Mix_Den),

while the other three were used as a control (C_Den). The NO_3^- concentration was measured prior to the addition of $^{15}\text{NO}_3^-$ and before the cores were mixed in order to calculate the $^{14}\text{N}/^{15}\text{N}$ ratio in the NO_3^- pool. Floating lids were then positioned on the top of all cores and the incubation was carried out, according to standard procedures (Dalsgaard *et al.*, 2000). At the end of the incubation, 10 mL of 7M ZnCl_2 were added to the water phase of all cores and then sediment and water were mixed. Part of the slurry was then transferred into 12.5 mL gas-tight vials; $^{14}\text{N}^{15}\text{N}$ and $^{15}\text{N}^{15}\text{N}$ abundance in N_2 was analysed by mass spectrometry at the National Environmental Research Agency (Silkeborg, Denmark). The rates of denitrification (D_{tot}) were calculated according to the equations and assumptions of Nielsen (1992): $D15 = p(^{15}\text{N}^{14}\text{N}) + 2p(^{15}\text{N}^{15}\text{N})$ and $D14 = p(^{15}\text{N}^{14}\text{N}) + 2p(^{14}\text{N}^{14}\text{N})$, where D15 and D14 = rates of denitrification based on $^{15}\text{NO}_3^-$ and $^{14}\text{NO}_3^-$, respectively; and $p(^{14}\text{N}^{14}\text{N})$, $p(^{15}\text{N}^{14}\text{N})$ and $p(^{15}\text{N}^{15}\text{N})$ = rates of production of labelled and unlabelled N_2 species. Because the $p(^{14}\text{N}^{14}\text{N})$ cannot be readily measured, estimation of D14 was obtained from: $D14 = D15 \times p(^{15}\text{N}^{14}\text{N}) / 2p(^{15}\text{N}^{15}\text{N})$.

RESULTS

Water column characterisation

In the period of the investigation, temperatures ranged between 3.0 and 23.5°C; pH was alkaline (7.83–7.98) and oxygen saturation was minimum in March (~61%) and maximum in December 2003 and February 2005 (~83%), with an average value of 76±10% (Tab. 1). DIN concentration peaked in winter (160–260 µM) and was minimum in summer (<25 µM in July 2004); NO_x^- was generally the dominant form of inorganic nitrogen, except in July 2004. SRP concentration was similar in March and July 2004 and peaked in December 2003 and February 2005. Chl *a* concentrations were low in summer (~6 µg L⁻¹ in July 2004) and relatively high in winter (21.8 µg L⁻¹ in December 2003).

Sediment characterisation and sedimentation rates

Surface sediments were fluffy and consisting of macrophyte fragments; the colour was light brown with

no apparent vertical discontinuities. The macrofauna community was poor with large individuals of the bivalve *Anodonta* sp. (not sampled in the cores used in this work) and a few chironomids. Many cores were discarded during sampling activities due to large bubbles breaking the sediment-water interface and resuspending particles; ebullition was evident also *in situ*.

Density, porosity, water and organic matter content and elemental composition were homogeneous between analysed slices along the 0–10 cm horizon at all sampling dates (ANOVA, $P > 0.05$). Sediment density (1.01–1.03 g cm⁻³) was close to that of water and not significantly different among seasons (ANOVA, $P > 0.05$); water content was always above 91% and sediment porosity above 0.86 (Tab. 2). Organic matter content tended to be higher in July and September 2004 (33.4±1.7 and 34.4±1.6%, respectively) and lower in December 2003 (31.0±1.4%), with an average value of 32.5±1.8% along the 0–10 cm horizon. Average values of organic C, total N and P were 17.43±0.88, 1.86±0.12 and 0.15±0.02% (w/w), respectively, and did not show seasonal trends (Tab. 2).

Gross sedimentation rates were similar on the two sampling dates when this process was measured, with average values of 37.7±7.0 and 44.8±5.4 g_{dw} m⁻² d⁻¹ calculated for March and September 2004, respectively.

Sediment profiles of oxygen, redox potential, pH, ammonium and soluble reactive phosphorous

Porewater O₂ and Eh values exhibited small scale sharp vertical variations. The thickness of the oxic sediment layer varied with seasons with highest values (~7 mm) measured in winter, and minimum (<2 mm) in summer (Fig. 2). Oxygen penetration was inversely correlated with both water temperature ($r^2 = 0.96$, $P < 0.001$) and organic matter content ($r^2 = 0.92$, $P < 0.001$). All profiles showed typical parabola-fitting depth trends indicating that the oxygen distribution was mostly governed by molecular diffusion between the oxic water and the oxygen-consuming sediment. None of the oxygen microprofiles showed subsurface peaks.

Water column Eh was always largely positive (>200 mV) with the exception of July (~50 mV). Sharp reduction in the Eh values was measured in all sampling dates within

Tab. 1. Physicochemical parameters of the water overlying sediments at the sampling station.

Date	T (°C)	pH	Conductivity (µS cm ⁻¹)	O ₂ (µM)	O ₂ (%)	NH ₄ ⁺ (µM)	NO _x ⁻ (µM)	SRP (µM)	Chl <i>a</i> (µg L ⁻¹)
12/03	3.0	7.86	409	334	83.1	9.8	149.1	0.36	21.8
03/04	8.0	7.91	497	222	60.8	6.2	78.6	0.09	9.9
07/04	23.0	7.98	545	206	74.8	15.1	8.5	0.09	5.9
09/04	23.5	7.83	303	208	78.9	7.0	60.0	0.00	8.8
02/05	4.0	7.95	639	319	83.3	15.3	245.0	0.40	13.3

SRP, soluble reactive phosphorous; Chl *a*, Chlorophyll *a*.

the upper 5 mm; below this horizon Eh was constant with depth and close to -300 mV. The redox potential discontinuity was quite overlapping the oxic-anoxic interface at all sampling dates with up to 500 mV drop (from +200 to -300 mV) measured in September 2004 (Fig. 2).

Porewater pH was generally slightly lower compared to that of the water column; maximum variations were measured in summer with a decrease of approximately 0.7 units in the 0-0.5 cm upper layer (from 7.8 to 7.1, July 2004). Minimum porewater pH values were slightly below 7 (6.7, July 2004); on the other dates values were rather constant and close to 7.5 (Fig. 2).

Porewater concentrations of NH_4^+ and SRP generally increased with depth at all sampling dates (Fig. 3). During the entire study, NH_4^+ concentrations in the upper 0-2 cm sediment layer were comparable to those measured in the water column (2-10 μM) and increased in the 2-3 cm layer; in the deepest investigated horizon (5-10 cm) ammonium levels were surprisingly high in December 2003 (554 \pm 246 μM). During 2004, NH_4^+ concentrations in the deepest slice peaked in July (386 \pm 119 μM), while minimum values were recorded in March and September (~100 and ~180 μM).

Despite the anoxic conditions and low Eh, porewater SRP concentrations were extremely low in the upper 5 cm horizon and very similar to those of the overlying water on four out of the five sampling dates, with values varying between 0.1 and 1.9 μM . The exception was December 2003 when SRP concentrations increased significantly with depth and peaked in the 5-10 cm slice (37 \pm 29 μM). Values measured in the deepest slice were close to 10 μM in March and July 2004 and below 5 μM in September 2004 and February 2005 (Fig. 3).

Oxygen, ammonium, and soluble reactive phosphorous benthic fluxes

Oxygen uptake by sediments was mostly regulated by water temperature, with highest rates determined in July (56.9 \pm 5.0 and 42.9 \pm 6.2 $\text{mmol m}^{-2} \text{d}^{-1}$ for total and diffusive uptake, respectively) and in September 2004 (56.9 \pm 23.7 and 48.1 \pm 10.3 $\text{mmol m}^{-2} \text{d}^{-1}$ for total and diffusive uptake,

respectively), and lowest rates measured in December 2003 (12.7 \pm 4.2 and 3.8 \pm 1.3 $\text{mmol m}^{-2} \text{d}^{-1}$ for total and diffusive uptake, respectively) (Fig. 4). The largest difference between measured and calculated oxygen fluxes was during winter months when the diffusive flux accounted for only 25 to 30% of the total sediment oxygen demand. On the contrary, in September and July 2004 diffusive uptake accounted for 75 to 84% of total oxygen uptake.

Sediments regenerated NH_4^+ to the water column on four out of the five sampling dates, with rates increasing from December 2003 (~200 $\mu\text{mol m}^{-2} \text{d}^{-1}$) to September 2004 (~5400 $\mu\text{mol m}^{-2} \text{d}^{-1}$). Only in February 2005 a net NH_4^+ uptake (-130 $\mu\text{mol m}^{-2} \text{d}^{-1}$) was measured (Fig. 5). As for oxygen demand, the seasonal evolution of NH_4^+ fluxes was temperature dependent. Instead, SRP fluxes were erratic and did not follow a clear seasonal pattern, increasing from December 2003 to July 2004 but shifting to negative in September 2004 (Fig. 5).

Calculated diffusive NH_4^+ and SRP fluxes were always positive and varied between 124 \pm 25 $\mu\text{mol m}^{-2} \text{d}^{-1}$ (March 2004) and 784 \pm 236 $\mu\text{mol m}^{-2} \text{d}^{-1}$ (July 2004) and between 1.4 (February 2005) and 17.5 $\mu\text{mol m}^{-2} \text{d}^{-1}$ (December 2003), respectively, without any seasonal trend.

Sediment resuspension, benthic fluxes and denitrification rates

Resuspension of surface sediments resulted in a sudden drop of oxygen and nitrate concentrations into the overlying water and in ammonium and reactive phosphorus release. Average oxygen concentration in the water column decreased from 0.29 to 0.07 mM, corresponding to an instantaneous O_2 consumption of 28.2 \pm 0.3 mmol m^{-2} . Similarly, nitrate concentration decreased from 34.5 to 10.5 μM , with a nitrate consumption of 3.13 \pm 0.43 mmol m^{-2} . Ammonium concentration in the water overlying sediments increased during resuspension from 11.0 to 63.7-103.0 μM , while water column SRP level increased from 0.20 to 0.25-0.83 μM . NH_4^+ and SRP release during sediment resuspension was equivalent to an efflux varying between 7.4 and 12.0 mmol m^{-2} and from 6.4 and 82.4 $\mu\text{mol m}^{-2}$, respectively.

Tab. 2. Main features of sediments at sampling station. As values of measured parameters were homogeneous in the considered vertical horizon (0-10 cm depth) data from the different layers were pooled. Average values \pm SD (n=15) are reported.

Date	Water content (%)	Density (g cm^{-3})	Porosity	Organic matter (%)	Organic carbon (%)	Total nitrogen (%)	Total phosphorus (%)
12/03	93.3 \pm 1.0	1.02 \pm 0.02	0.95 \pm 0.01	31.0 \pm 1.4	17.18 \pm 1.03	1.79 \pm 0.08	0.13 \pm 0.01
03/04	93.7 \pm 1.2	1.01 \pm 0.04	0.95 \pm 0.03	31.8 \pm 1.2	17.20 \pm 0.82	1.87 \pm 0.07	0.17 \pm 0.01
07/04	92.5 \pm 1.4	1.03 \pm 0.11	0.95 \pm 0.09	33.4 \pm 1.7	17.25 \pm 0.84	1.84 \pm 0.09	0.14 \pm 0.01
09/04	93.3 \pm 1.0	1.02 \pm 0.02	0.95 \pm 0.02	34.4 \pm 1.6	17.86 \pm 0.65	1.88 \pm 0.21	0.14 \pm 0.01
02/05	92.4 \pm 1.0	1.01 \pm 0.02	0.91 \pm 0.02	32.1 \pm 1.0	17.64 \pm 0.94	1.92 \pm 0.08	0.16 \pm 0.01

Sediment-water fluxes of O_2 , NH_4^+ and SRP measured after sediment mixing and renewal of water overlying sediments were not statistically different from those measured in control cores (ANOVA, $P>0.05$) and comparable with those found in September (Fig. 6). There was just a tendency towards decreasing rates of oxygen uptake and ammonium release in mixed compared to control cores. Nitrate fluxes were negative in

both control and mixed cores, with a tendency towards higher consumption rates in control cores (-1.78 to -14.96 $mmol\ m^{-2}\ d^{-1}$) than in mixed cores (-1.66 to -3.54 $mmol\ m^{-2}\ d^{-1}$).

Denitrification rates measured in resuspended sediments (12.40 - 19.80 $mmol\ m^{-2}\ d^{-1}$) were significantly higher (ANOVA, $P<0.05$) than rates measured in control cores (0.33 - 1.34 $mmol\ m^{-2}\ d^{-1}$) (Fig. 6).

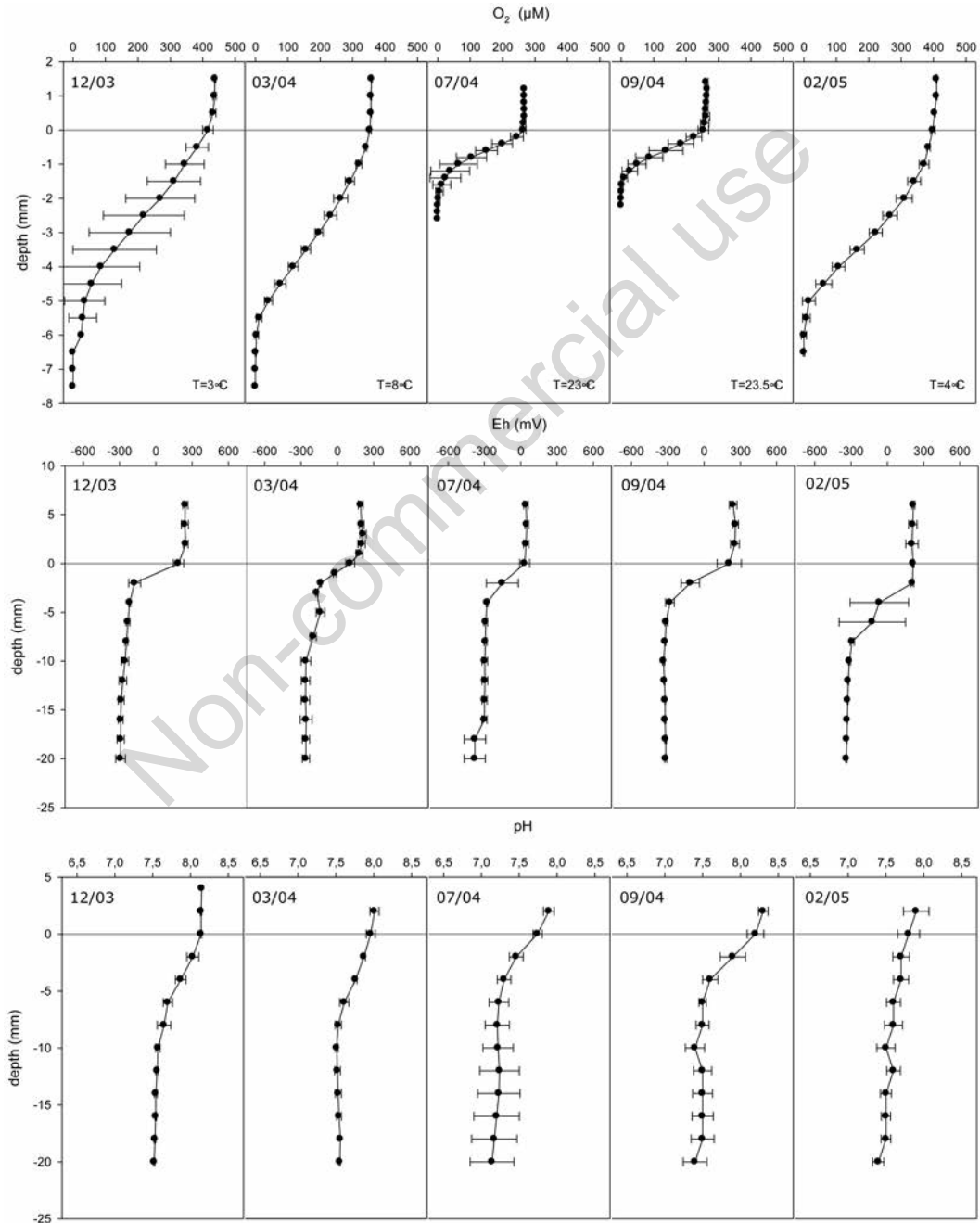


Fig. 2. Seasonal sediment profiles of dissolved oxygen, pH and Eh measured at sampling station via micro and mini sensors. Average values \pm SD ($n=3$) are reported.

DISCUSSION

Vertical profiles in unconsolidated sediments

Gross sedimentation at sampling station is comparable to that reported for shallow lakes exhibiting high rates of sediment resuspension (Niemisto *et al.*, 2008; Nurminen and Horppila, 2009). Sediment resuspension is recognised as an important physical process affecting a number of ecological aspects of shallow water bodies (Evans, 1994; Scheffer, 1998) and accounts for 40-90% of total sedimentation rates (Jimenez-Montealegre *et al.*, 2002; Niemisto *et al.*, 2008). If this is true also at our sampling

site, net sedimentation rate at the study area can be estimated to be 4-25 $\text{g}_{\text{dw}} \text{m}^{-2} \text{d}^{-1}$, which is equivalent to a deposition of fresh organic matter of 2-13 $\text{cm} \text{y}^{-1}$. This ultimately means that the sediment layer characterised in the present study was very recent.

In the shallow study area, sediment density along the upper 0-10 cm horizon was very close to that of water. Winds or benthic fish such as carps, abundant in the study area, can easily resuspend, mix and homogenise settled particles (Sondergaard *et al.*, 1992; Breukelaar *et al.*, 1994; Cline *et al.*, 1994; Mayer *et al.*, 1999; Persson and Svensson, 2006). An evidence of this phenomenon is given by

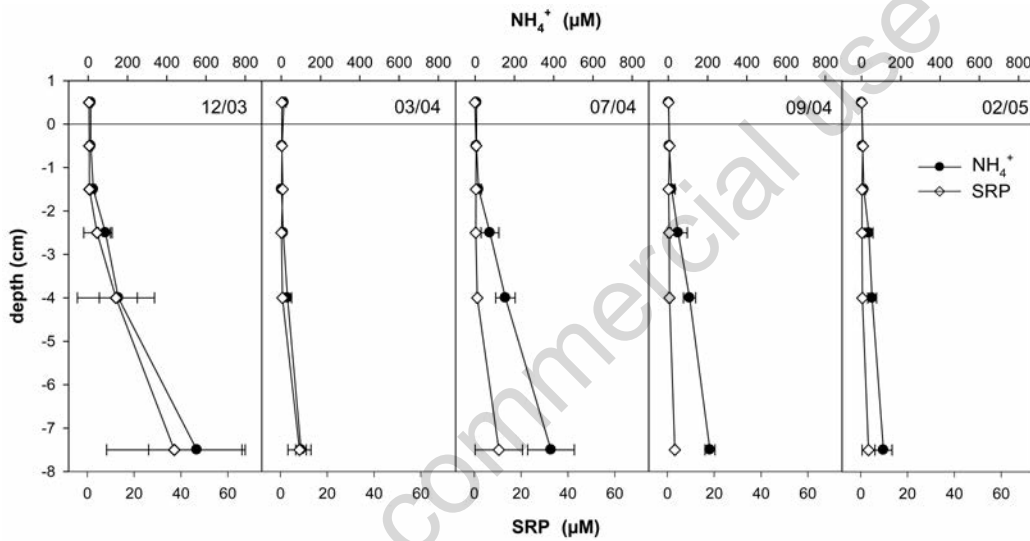


Fig. 3. Depth profiles of ammonium (NH_4^+) and soluble reactive phosphorous (SRP) concentration within sediments; results from the whole investigation cycle are shown. For each profile average values \pm SD ($n=3$) are reported.

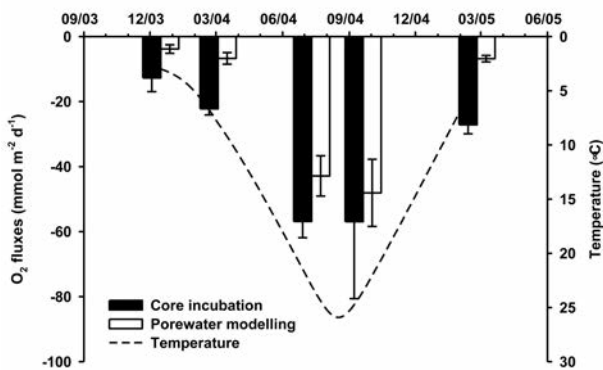


Fig. 4. Dark sediment oxygen demand measured via intact core incubation and calculated from porewater profiles. Average values \pm SD ($n=3$) are reported.

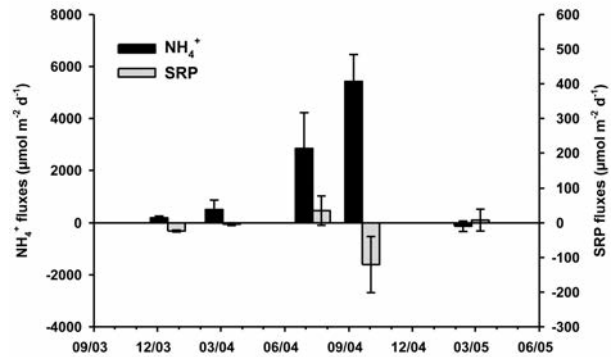


Fig. 5. Ammonium (NH_4^+) and soluble reactive phosphorous (SRP) exchanges across the sediment-water interface measured via intact core incubation. Average values \pm SD ($n=3$) are reported.

most of the sediment profiles, which were homogeneous along the investigated horizon in all sampling periods.

Oxygen penetration at the sampling site was limited to a few millimetres depth and it was regulated by water temperature and bacterial activity. Below the oxic layer, strictly anaerobic metabolism probably driven by methanogenesis was coupled to very low Eh values; methanogenesis and gas ebullition contributes to sediment mixing (Segers, 1998; Liikanen *et al.*, 2003; Longhi *et al.*, 2008).

Benthic fluxes in unconsolidated sediments

Total oxygen uptake rates were comparable to those reported for other organic-rich freshwater sediments (Sweerts *et al.*, 1991b; Fisher and Reddy, 2001). Oxygen uptake in whole core incubation exceeded that calculated from microprofiles as it includes respiration by plankton, bacteria, meio- and macrofauna, while diffusive fluxes account for microbial respiration alone (van Der Loeff *et al.*, 1984; Aller, 1988; Archer and Devol, 1992; Glud *et al.*, 1994). Maximum differences between oxygen fluxes from core incubations and microprofiling were calculated for winter months when macrofauna was limited to a few chironomids larvae. No information is available for meiofauna abundance in these sediments and meiofauna has the potential for high respiration rates (Lasserre *et al.*, 1976).

Besides the presence of biota, several other factors could influence oxygen flux measurements in core incubation. For example, oxygen respiration in core incubations can overestimate *in situ* oxygen demand when stirring is too vigorous (Fisher and Reddy, 2001) and this could be the case at our sampling station, where water is generally stagnant. We suggest that rates measured via core incubations and calculated from micro profiles mod-

elling set the upper and lower limits of the true oxygen demand for these sediments, respectively.

On an annual basis, the net exchange of NH_4^+ and SRP across the sediment-water interface was estimated in $\sim 750 \text{ mmol NH}_4^+ \text{ m}^{-2} \text{ y}^{-1}$ and $\sim 9 \text{ mmol SRP m}^{-2} \text{ y}^{-1}$, suggesting that Busatello sediments were a net N source and a net P sink.

The flux of NH_4^+ calculated from porewater modelling was approximately 5 times lower than that calculated from core incubation and did not follow a seasonal pattern. This is an unexpected result since mineralisation rates and accumulation of solutes in interstitial waters are generally correlated to water temperature (Rysgaard *et al.*, 1995; Clavero *et al.*, 2000). The failure of diffusive fluxes to mirror expected seasonal patterns can be a consequence of porewater profiles vertical resolution. The resolution we adopted for sediment slices close to the interface (1 cm) is probably not accurate enough to record all the processes occurring at the sediment surface (Urban *et al.*, 1997), but it was a consequence of sediments at sampling site, more comparable to an aqueous slurry than to a hard bottom. Besides spatial resolution, several other factors could influence the interpretation of porewater profiles. The fitting methodology of porewater profiles assumes that molecular diffusion is the most important transport process in the sediments and that steady-state conditions are present; but within the considered fluffy sediments the steady-state of porewater solutes is probably difficult to establish. We hypothesise that resuspension events may occur frequently enough to keep the sediments constantly mixed, in particular during summer months when faunal activity peak and ebullition phenomena are frequent. This hypothesis is in agreement with the lack of seasonal trends in nutrient porewater profiles and could explain differences between measured and calculated nutrient fluxes. This was not the same in the case of oxygen as oxygen penetration decreased by a factor 3 from winter to summer. It is likely that, after events of sediment resuspension, oxygen profiles reach a steady-state in a very short time lag, due to its rapid consumption in deep horizon (Gerhardt and Schink, 2005). The situation is probably different for solutes like NH_4^+ and SRP, which are end products of mineralisation processes and enter into a number of both microbially-mediated and geochemical reactions. For these solutes the establishment of steady-state sedimentary profiles can require a longer time lag as it could be susceptible to factors like the vertical re-distribution of fresh and old organic matter, the development of specialised, slow-growing bacterial communities (*i.e.*, nitrifiers) or the regeneration of biogeochemical buffers (*i.e.*, Fe^{3+} pools). This could also explain the discrepancies observed between NH_4^+ and SRP fluxes measured via incubation and calculated from porewater concentrations. In this organic rich bottom, SRP dynamics are probably influenced by adsorption processes to dissolved organic molecules which make interpretation

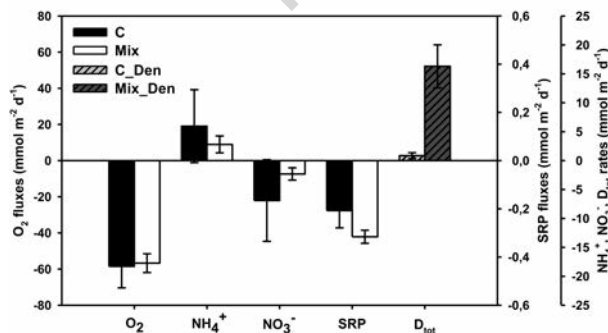


Fig. 6. Oxygen (O_2), ammonium (NH_4^+), nitrate (NO_3^-) and soluble reactive phosphorous (SRP) fluxes across the sediment-water interface and denitrification rates (D_{tot}) measured in control cores (C and C_Den) and in cores where sediments were resuspended (Mix and Mix_Den, see text for major details). Average values \pm SD ($n=3$) are reported.

of profiles complicated (Reddy *et al.*, 1999). To conclude, we believe that nutrient fluxes calculated from porewater profiles are not reliable due to insufficient vertical resolution close to the interface and absence of steady-state gradients between the water column and the porewater from the upper sediment layers.

Is sediment resuspension relevant for benthic fluxes and denitrification rates?

Results from this experiment indicate: i) an instantaneous release of NH_4^+ and SRP from porewater to the water column and a rapid consumption of O_2 and NO_3^- ; ii) that resuspension and mixing of recalcitrant macrophytic litter did not affect significantly benthic fluxes and iii) that denitrification rates underwent a significant transient increase as a consequence of sediment resuspension. Ammonium and reactive phosphorus release following sediment mixing averaged $9.0 \pm 2.6 \text{ mmol m}^{-2}$ and $41.6 \pm 38.3 \text{ } \mu\text{mol m}^{-2}$, respectively, and were approximately 20 and 16 times higher than daily fluxes measured via core incubations, respectively. This means that nutrient release during a single resuspension event was equivalent to the regeneration of N and P occurring in 2-3 weeks. However, sediment-water fluxes of NH_4^+ were roughly counterbalanced by nitrate consumption rates, indicating that nitrogen production and consumption processes were in equilibrium.

The instantaneous decrease in water column oxygen and nitrate concentrations due to resuspension is partly due to dilution of overlying oxygenated water with anoxic porewater, and partly due to oxidation of reduced end products of mineralisation. These amounts were calculated knowing oxygen and nitrate concentrations in water column before and immediately after sediment mixing and the total volume of porewater. About 65 and 71% of oxygen and nitrate decrease was due to dilution, respectively, while about 35 and 29% was due to chemical consumption. If we assume the occurrence of one resuspension event per day, the overall oxygen consumption during the event accounts for 55% of total daily oxygen demand by sediments, as measured with intact cores incubations. This result is in agreement with those found by Adams *et al.* (1982) and Sweerts *et al.* (1991a), which have estimated the oxygen equivalents necessary for the oxidation of reduced inorganic compounds (NH_4^+ , Fe^{2+} , Mn^{2+} , HS^- and CH_4) in lake sediments. Adams *et al.* (1982) calculated that the oxidation of these compounds at the sediment-water interface was equivalent to 35% of the sediment oxygen demand, while Sweerts *et al.* (1991a) calculated that the oxidation of anoxic mineralisation end products accounted for up to 70% of the sediment oxygen consumption.

The significant stimulation of D_{tot} by resuspension is a consequence of NO_3^- transport within reduced pore-

waters. When resuspension occurs, the NO_3^- (and O_2) in the overlying water are transported in relatively deep sediment horizons where they are not generally present. Here, nitrate can be rapidly converted into N_2 or NH_4^+ , while oxygen can stimulate nitrification-coupled denitrification (Rysgaard *et al.*, 1995). After the exhaustion of the supplied NO_3^- (and O_2) we expect denitrification rates to be again regulated by the diffusion of nitrate from the water column and by nitrification within sediments and thus similar to those measured prior to sediment resuspension.

The whole effect of all these simultaneous processes – occurring in a relatively short time lag – is probably to enhance the rates of organic matter mineralisation.

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