

# Diatom community biodiversity in an Alpine protected area: a study in the Maritime Alps Natural Park

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## ABSTRACT

As part of the "All Taxa Biodiversity Inventories" (ATBIs) coordinated by the European Distributed Institute of Taxonomy (EDIT), we analysed diatom communities colonizing different habitats of the Valasco Valley (Maritime Alps Natural Park). The aim of this research was to shed light on the diatom richness in an Alpine context through i) the collection of data concerning diatom communities inhabiting an unexplored watershed of the Maritime Alps Natural Park, including all of the most important aquatic habitats (comparison among river, springs and peat bogs) and assemblages (epilithic and epiphytic diatom communities); ii) analysis of the main environmental factors driving the development of diatom communities in different habitats. We completed a list of 174 diatom taxa. In general, river samples were poorer in terms of species richness than spring and peat bog ones, probably due to the selective role of the fast flow, while springs sheltered the highest biodiversity of the Valasco Valley. Peat bog communities were mainly composed of acidophilous taxa, sometimes planktonic and forming colonies. Epilithic and epiphytic samples did not show significant differences in terms of composition and biodiversity, even though it was statistically possible to identify indicator species for each assemblage. The study also highlighted the presence of several taxa included in the German Red List as endangered or decreasing, especially in the epiphytic samples. Since the water nutrient level and substrate geology were similar among habitats, the main environmental factors shaping Valasco diatom communities were water velocity and pH.

Key words: Valasco Valley, biodiversity, epilithic, epiphytic, springs, peat bogs

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## 1. INTRODUCTION

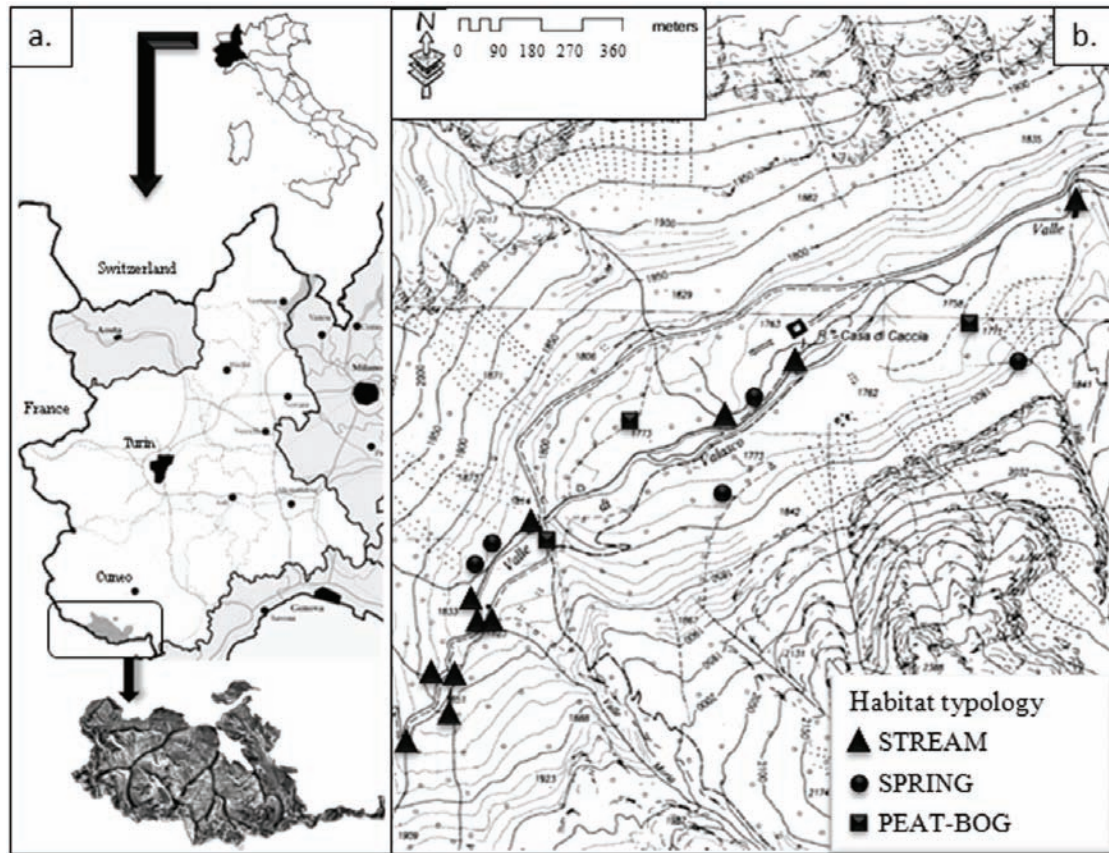
Despite the effort to significantly reduce biodiversity loss by 2010, recent research has confirmed the extinction of several plant and animal species listed as endangered in the IUCN Red Lists, while allochthonous taxa are increasing globally (Butchart *et al.* 2010). Therefore, fragile ecosystems such as Alpine streams should be carefully monitored, as they are exposed to extreme conditions due to their harsh nature (Cantonati *et al.* 2001; Cantonati & Spitale 2009) and human exploitation (Fránková *et al.* 2009). For this reason, the analysis of biological assemblages inhabiting protected areas is a useful tool to establish reference and autochthonous communities not affected by human pressure and to identify endangered species that should be protected (Werum 2001; Fránková *et al.* 2009).

Benthic diatoms are regularly used as biological indicators for the environmental assessment of river water quality (included in the Water Framework Directive 2000/60). The analysis of diatom communities is a tool to guarantee an ecological and sustainable use of the water resources and the correct elaboration of guidelines for their preservation. In Italy, studies analysing diatom assemblages in high mountain habitats mainly cover the Eastern Alps (Cantonati 1998a, 1998b, 1999; Cantonati & Ortler 1998; Cantonati & Pipp 2000; Cantonati 2001; Cantonati *et al.* 2001; Cantonati & Lange-Bertalot 2006; Cantonati *et al.* 2006; Cantonati *et al.* 2007; Cantonati & Spitale 2009), while data on the Western Alps are more recent (Battagazzore *et al.* 2004; Falasco *et al.* 2007; Bona *et al.* 2008; Falasco *et al.* 2008; Mobili *et al.* 2008; Falasco *et al.* 2009a; Bona *et al.* 2011).

High mountain streams are extremely oligotrophic ecosystems where the geology of the substrate represents an important environmental factor in shaping diatom communities. On the one hand, siliceous substrates shelter the highest biodiversity, with indicator species characteristic of low Ca water content. At the local scale, current velocity and pH are the most important variables influencing diatom community composition since the geology is the same among habitats (Sabater & Roca 1992; Cantonati & Spitale 2009). On the other hand, the importance of the substrate typology (lithic or plants) is still poorly understood and controversial: while Bertrand *et al.* (2004) showed no significant differences between communities developed on cobbles or macrophytes, Fránková *et al.* (2009) hypothesized a specific relationship between *Sphagnum* and diatoms. Nevertheless, studies on this topic are still rare.

In high mountains, springs usually shelter high biodiversity due to low variability of the environmental factors over the seasons. Because of their low nutrient content, moderate water flow and constant temperature, springs can be considered suitable habitats for a wide range of species (Cantonati 1998a, 1998b, 1999). Indeed, springs offer protection for endangered and rare

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**Fig. 1.** a. Maritime Alps Natural Park location; b. Sampling site locations and relative habitat type.

taxa in addition to the common oligosaprobous ones (Cantonati 2001; Cantonati & Spitale 2009). At present, the only Red List available for evaluating the conservation status of diatom species is the German one (Lange-Bertalot & Steindorf 1996): there are few historical literature data on diatom distribution and conservation status from other European countries.

This research, carried out as part of the "All Taxa Biodiversity Inventories" (ATBI), was aimed at the elaboration of plant and animal check-lists for the Maritime Alps Natural Park. We examined three different habitat types in the Valasco Valley (the main Valasco River, some rheocrene springs and peat bogs), collecting samples from stones and submerged plants (epilithic and epiphytic samples). The comparison with the German Red List provided information on the presence of rare or endangered species. In addition to the production of a complete list of diatom taxa colonizing the Valasco Valley, this research aims to provide important information concerning the ecological preferences of diatom species in relation to the most important environmental variables.

## 2. METHODS

The Italian Maritime Alps Natural Park (Fig. 1a) is located in the southernmost section of the Alpine chain close to the French border. It has been twinned with the

French Mercantour National Park since 1987 but it was officially established only in 1995. Due to its proximity to the Ligurian Sea and the presence of high peaks (over 3000 m a.s.l.), the Maritime Alps Natural Park offers a heterogeneous habitat for animals and plants, with very high biodiversity.

The present study was conducted in the Valasco Valley (1760–1832 m a.s.l.). The principal watercourse is the Valasco River, flowing in the valley bottom where there are also many small springs and peat bogs.

In summer 2009, we sampled 19 sites representing different habitat types: 11 sites in the main river, 5 in small rheocrene springs and 3 in peat bogs (Fig. 1b). Permanently wet springs and peat bogs were chosen. At each sampling site, we evaluated the main physical and chemical parameters by means of a multiparametric probe (HYDROLAB QUANTA): conductivity, dissolved oxygen, pH and temperature. We recorded the water velocity in three different sections of each transect with a current meter (HYDRO-BIOS KIEL). Water quality parameters (N-NH<sub>4</sub>, N-NO<sub>3</sub>, P-PO<sub>4</sub>, BOD<sub>5</sub>, Ca and water hardness) were analysed by means of a Dr Lange LASA 100 spectrophotometer with analysis kits (LCK 304; LCK 554; LCK 327; LCK 339; LCK 349).

When possible, we collected diatom communities from different substrates (31 samples from 19 sites). We sampled the epilithon by randomly choosing at least five

**Tab. 1.** Physical and chemical parameters recorded in the field and analysed in the laboratory. R=river, S=springs and P=peat-bogs; EL=epilithic sample, EP=epiphytic sample. COND=conductivity; T=temperature; V=water velocity (mean values); HARD=hardness. \* below the instrument detection limit.

Site Code	Site Type	Sample Type		Cond. ( $\mu\text{S cm}^{-1}$ )	O <sub>2</sub> (mg L <sup>-1</sup> )	O <sub>2</sub> (%)	pH	T (°C)	V (m s <sup>-1</sup> )	BOD <sub>5</sub> (mg L <sup>-1</sup> )	Ca (mg L <sup>-1</sup> )	Hard (°dH)	N-NH <sub>4</sub> (mg L <sup>-1</sup> )	N-NO <sub>3</sub> (mg L <sup>-1</sup> )	P-PO <sub>4</sub> (mg L <sup>-1</sup> )
		EL	EP												
9042	R	+	+	19	9.85	87.8	7.59	8.00	0.52	1.23	5.73	0.80	0.020	0.237	0.044
9044	R	+	+	21	9.70	81.5	7.00	8.00	0.26	0.00	5.63	0.79	0.020	0.283	0.011
9045	R	+	+	23	10.25	81.2	7.00	5.55	0.97	1.31	7.30	1.02	0.028	0.063	0.007
9046	R	+		24	10.52	84.9	7.35	6.66	0.32	0.20	5.74	0.80	0.018	0.313	0.014
9047	R	+		14	8.33	79.3	6.99	13.27	0.37	0.90	5.55	0.78	0.024	0.197	0.0005*
9048	R	+		19	9.45	77.7	7.02	7.04	0.39	0.64	7.72	1.08	0.024	0.119	0.008
9049	R	+		20	8.74	77.3	6.95	9.78	0.46	0.78	7.03	0.99	0.047	0.260	0.015
9050	R	+	+	16	7.39	73.1	6.66	14.98	0.09	1.00	6.31	0.88	0.031	0.148	0.003
9054	R	+		20	9.32	81.5	6.81	9.69	0.67	4.43	6.59	0.93	0.028	0.236	0.006
9056	R	+		15	9.97	97.3	6.80	12.99	0.51	0.44	5.08	0.71	0.030	0.080	0.010
9058	R	+	+	20	12.73	110.0	6.64	7.88	0.60	1.66	9.62	1.35	0.022	0.311	0.003
9051	S	+	+	19	9.40	77.3	6.75	7.4	0.05*	4.29	6.14	0.86	0.039	0.155	0.005
9052	S	+	+	18	9.14	75.9	6.30	7.29	0.05*	2.56	8.04	1.13	0.026	0.271	0.006
9055	S	+	+	16	8.22	70.3	6.03	8.6	0.05*	0.99	4.94	0.69	0.029	0.164	0.010
9059	S	+		13	6.58	104.0	6.48	6.48	0.05*	4.79	4.08	0.57	0.044	0.108	0.008
9061	S	+	+	27	12.30	98.9	6.37	5.34	0.05*	1.78	7.57	1.06	0.038	0.816	0.001
9053	P	+	+	26	9.30	75.5	6.63	6.81	0.05*	0.91	5.96	0.88	0.032	0.160	0.0005*
9057	P	+	+	17	9.38	96.2	6.68	15.3	0.05*	1.30	6.41	0.90	0.049	0.207	0.012
9060	P	+	+	21	10.00	95.2	6.17	12.01	0.05*	7.92	7.85	1.10	0.025	0.268	0.0005*

cobbles along each sampling transect (Kelly *et al.* 1998). The epiphytic communities were collected from the dominant macrophytes along the transect, including bryophytes and entire living plants, by squeezing and cutting entire portions of them (Cantonati *et al.* 2007). In order to exclude moisture content variability, we chose only completely submerged substrates. All 31 samples were treated with hydrogen peroxide (100 vol.) and HCl (1N) (EN 13946, 2003). Glass slides were mounted with Naphrax<sup>®</sup> and analysed under a Leitz Diaplan microscope with a 100 $\times$  immersion objective. Diatoms were identified mainly according to Krammer & Lange-Bertalot (1986-1991a, b), Lange-Bertalot & Metzeltin (1996), Krammer (1997a, b), Reichardt (1999), Lange-Bertalot (2001), Krammer (2000; 2002; 2003) and Werum & Lange-Bertalot (2004). In a first stage of the analysis, we identified all the species on the whole glass slides in order to complete a check-list of the recorded species. Later, we defined the relative abundances by identifying at least 400 valves per sample.

Significant differences in physical and chemical parameters and biological indices (IPS, TID and diversity indices) among habitat and substrate types were highlighted with the non-parametric Kruskal-Wallis test (SPSS Statistics 17.0). Multivariate analyses were carried out with PC-ORD software (McCune & Mefford 1999). In particular, Canonical Correspondence Analysis (CCA), Indicator Species Analysis (ISA), Multiresponse Permutation Procedure (MRPP) and Non-metric Multidimensional Scaling (NMS) were performed on biological communities, while Principal Component Analysis (PCA) was applied to the environmental dataset. ISA was used to define significantly characteristic species of habitats (river, peat bogs and springs) and assemblages (epilithic and epiphytic samples). NMS of

diatom assemblages was used to test similarities and differences in diatom assemblages sampled in different habitat types and on different substrates. We set the NMS by choosing a random starting configuration: 40 runs with real data and SORESENSE (Bray-Curtis) as the distance measure. MRPP was used to test the hypothesis of no differences between the groups obtained from the NMS analysis. PCA was used to reduce the environmental dataset to a synthetic matrix to be used in the CCA analysis. The latter analysis was performed to better understand the relationships among environmental parameters and diatom communities.

### 3. RESULTS

#### 3.1. Physical and chemical parameters

Physical and chemical parameters detected in the water column are shown in table 1. All the sampling sites were located on a siliceous substrate; indeed, the Ca concentrations were very low (below 10 mg L<sup>-1</sup>). In general, no differences were found among the three sampled habitat types in terms of nutrient content and conductivity: all the sites could be classified as oligotrophic, with low electrolyte content and low organic loading. The highest value of BOD<sub>5</sub> was detected in a peat bog (site 9060). As expected, the current velocity was significantly higher in the main river than in springs or peat bogs, where the flow was below the instrument detection limit. Moreover, the pH was significantly lower in peat bogs than in the other habitat types ( $p < 0.01$ ; river>springs>peat bogs).

#### 3.2. Diatom communities

In total, 174 taxa belonging to 41 genera were recognized in the samples (see Appendix 1). In general,

**Tab. 2.** Typical assemblages of the three habitat types. Taxa were selected according to their abundance (>3% in at least one sample) and frequency (recorded in at least 25% of the samples belonging to a certain habitat type). Taxa are listed in order of abundance.

RIVER	<i>Achnanthydium minutissimum</i> , <i>Diatoma mesodon</i> , <i>Fragilaria arcus</i> , <i>Achnanthydium lineare</i> , <i>Fragilaria rumpens</i> , <i>Diatoma hyemalis</i> , <i>Diatoma</i> sp., <i>Gomphonema parvulum</i> var. <i>exilissimum</i> , <i>Planothidium haynaldii</i> , <i>Achnanthydium pyrenaicum</i> , <i>Encyonema silesiacum</i> , <i>Gomphonema pseudotenellum</i> , <i>Gomphonema micropus</i> , <i>Planothidium lanceolatum</i> , <i>Encyonopsis minuta</i> , <i>Gomphonema pumilum</i> var. <i>elegans</i> , <i>Nitzschia inconspicua</i> , <i>Neofragilaria virescens</i> , <i>Cymbella hantzschiana</i> var. <i>hantzschiana</i>
SPRING	<i>Diatoma mesodon</i> , <i>Achnanthydium minutissimum</i> , <i>Neofragilaria virescens</i> , <i>Achnanthes daonensis</i> , <i>Psammothidium oblongellum</i> , <i>Fragilaria rumpens</i> , <i>Diatoma</i> sp., <i>Aulacoseira alpigena</i> , <i>Gomphonema parvulum</i> var. <i>exilissimum</i> , <i>Encyonema silesiacum</i> , <i>Fragilaria arcus</i> , <i>Brachysira brebissonii</i> , <i>Eunotia minor</i> , <i>Eunotia exigua</i> var. <i>tenella</i> , <i>Staurorsira venter</i> , <i>Achnanthydium pyrenaicum</i> , <i>Achnanthydium lineare</i> , <i>Meridion circulare</i> , <i>Stauriforma exiguiiformis</i> , <i>Diademesis perpusilla</i> , <i>Aulacoseira paffiana</i>
PEAT-BOGS	<i>Achnanthydium minutissimum</i> , <i>Tabellaria flocculosa</i> , <i>Achnanthes daonensis</i> , <i>Aulacoseira alpigena</i> , <i>Fragilaria rumpens</i> , <i>Staurorsira venter</i> , <i>Psammothidium oblongellum</i> , <i>Eunotia exigua</i> var. <i>tenella</i> , <i>Diatoma mesodon</i> , <i>Brachysira brebissonii</i> , <i>Diatoma</i> sp., <i>Gomphonema parvulum</i> var. <i>exilissimum</i> , <i>Achnanthydium lineare</i> , <i>Neofragilaria virescens</i> , <i>Aulacoseira paffiana</i>

**Tab. 3.** Number of recorded species in the whole glass slide and in the inventories; main diatom and diversity indices. R=river, S=springs and P=peat-bogs; EL=epilithic sample, EP=epiphytic sample. N tot= number of species identified in the whole glass slide; N inv= number of species identified during the inventories (out of 400 individuals); B-P=Berger-Parker; J= Equitability; Ev= Evenness; Men= Menhinick; H= Shannon; I-D= Simpson.

Code Site	Type	N tot	N inv	Diatom Indices		Diversity Indices					
				IPS/20	TID/4	B-P	J	Ev	Men	H	I-D
9042	R-EL	37	17	19.2	1.09	0.54	0.54	0.54	0.97	2.41	0.66
9042	R-EP	31	24	19.6	1.00	0.59	0.52	0.52	1.26	2.48	0.63
9044	R-EL	37	19	19.6	1.00	0.41	0.66	0.65	0.94	2.87	0.78
9044	R-EP	38	28	19.5	0.97	0.46	0.56	0.56	1.43	2.74	0.73
9045	R-EL	17	14	19.9	0.86	0.31	0.62	0.62	0.75	2.48	0.77
9045	R-EP	29	17	19.0	0.78	0.53	0.52	0.52	0.94	2.29	0.66
9046	R-EL	23	21	19.7	0.88	0.28	0.73	0.73	1.11	3.30	0.85
9047	R-EL	21	18	17.1	2.18	0.40	0.62	0.62	0.96	2.72	0.77
9048	R-EL	16	15	19.8	0.95	0.53	0.52	0.52	0.79	2.14	0.65
9049	R-EL	19	15	19.6	0.82	0.40	0.67	0.67	0.83	2.83	0.78
9050	R-EL	30	22	19.7	1.03	0.68	0.42	0.42	1.15	1.91	0.52
9050	R-EP	31	21	19.5	1.05	0.55	0.56	0.56	1.09	2.52	0.66
9054	R-EL	18	14	19.9	0.88	0.51	0.50	0.50	0.74	2.00	0.64
9056	R-EL	18	14	19.9	0.96	0.49	0.48	0.48	0.74	1.87	0.65
9058	R-EL	17	14	19.7	0.98	0.57	0.51	0.51	0.77	2.08	0.63
9058	R-EP	31	18	19.5	0.96	0.25	0.73	0.73	0.98	3.20	0.85
9051	S-EL	37	19	17.7	1.11	0.32	0.70	0.70	1.23	3.50	0.85
9051	S-EP	29	18	16.7	1.12	0.29	0.71	0.71	1.07	3.28	0.84
9052	S-EL	32	24	19.6	0.86	0.35	0.72	0.72	1.29	3.45	0.84
9052	S-EP	31	25	18.6	0.92	0.29	0.66	0.66	1.36	3.20	0.83
9055	S-EL	45	38	18.6	1.13	0.15	0.83	0.82	1.95	4.39	0.93
9055	S-EP	30	18	19.4	0.94	0.40	0.59	0.59	1.03	2.67	0.75
9059	S-EL	29	29	18.7	1.00	0.18	0.73	0.73	1.44	3.53	0.88
9061	S-EL	31	24	18.7	0.91	0.24	0.68	0.68	1.27	3.25	0.85
9061	S-EP	18	17	19.1	0.94	0.27	0.77	0.77	1.07	3.13	0.84
9053	P-EL	24	18	17.4	1.28	0.24	0.75	0.75	0.94	3.23	0.85
9053	P-EP	34	22	17.4	1.51	0.20	0.73	0.73	1.27	3.55	0.88
9057	P-EL	29	22	19.2	1.01	0.44	0.61	0.61	1.18	2.81	0.75
9057	P-EP	42	31	18.9	1.05	0.38	0.66	0.66	1.64	3.38	0.81
9060	P-EL	36	25	18.7	0.92	0.35	0.68	0.68	1.27	3.21	0.83
9060	P-EP	37	18	18.9	0.86	0.57	0.56	0.56	1.07	2.59	0.65

communities were composed of xeno-oligosaprobous taxa, characteristic of high-altitude streams with low Ca and electrolyte content. The most frequent taxa were *Achnanthydium minutissimum* s.l. (detected in all the samples), *Achnanthydium daonense*, *Diatoma mesodon*, *Encyonema silesiacum*, *Fragilaria rumpens*, *Gomphonema parvulum* var. *exilissimum* (recorded in at least 87% of the samples).

The numbers of taxa recorded in the three habitat types were significantly different ( $p < 0.01$ ): springs > peat bogs > river. Spring communities presented the highest number of taxa: 100 species in 5 samples. River communities were generally composed of circumneutral

(ca 70%) and rheophilous taxa such as *Achnanthydium lineare*, *Diatoma hyemalis* (ISA;  $p < 0.05$ ), *D. mesodon* and *Fragilaria arcus* (ISA;  $p < 0.01$ ). Peat bogs had the lowest flow velocity and sheltered colonial or filamentous species such as *Aulacoseira alpigena*, *Aulacoseira paffiana* (ISA;  $p < 0.5$ ), *F. rumpens* and *Tabellaria flocculosa* (ISA;  $p < 0.5$ ), with a high percentage of acidophilous (ca 32%) and acidobiontic (ca 0.5%) taxa. Characterizing assemblages for each habitat type are proposed in table 2.

The biodiversity of diatom communities was particularly high (Tab. 3); we recorded significant differences in all the diversity indices calculated for the three

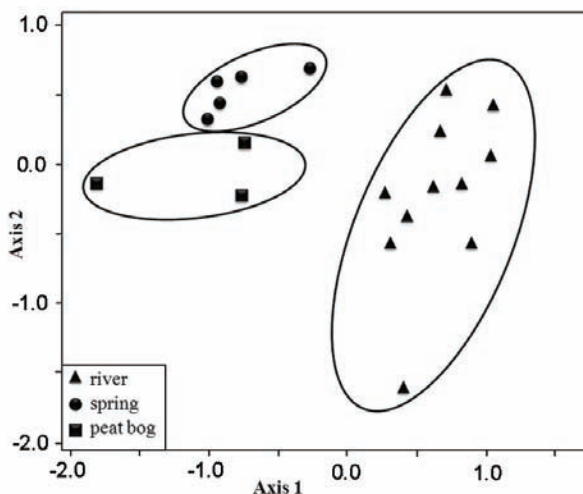
habitats: Berger-Parker ( $p < 0.01$ ), Equitability ( $p < 0.01$ ), Shannon ( $p < 0.001$ ), Simpson ( $p < 0.01$ ). The highest biodiversity values were obtained in springs.

The sampled sites offered suitable habitats for species of conservation interest. Indeed, about 30% of the recorded taxa were included in the Red List as "decreasing" or "endangered" (Lange-Bertalot & Steindorf 1996). Peat bogs seem to provide the best conditions for species threatened with extinction (*Naviculadicta detenta*), severely endangered species (*Eunotia botuliformis*, *Eunotia steineckii* and *Eunotia tetraodon*) and endangered ones (*Amphora inariensis*, *Cavinula pseudoscutiformis*, *Chamaepinnularia soehrensii* var. *hasiaca*, *Navicula angusta*, *Neidium alpinum*, *Neidium bisulcatum* and *Stauroforma exiguiiformis*). The highest percentage of taxa included in the Red List was found in epiphytic samples (35% vs 28% epilithic ones).

There were no significant differences in the number of taxa or the biodiversity index values among epilithic and epiphytic assemblages. Nevertheless, nearly 17% of the detected species were exclusively found on plant substrates. The indicator species for epiphytic assemblages was *Neofragilaria virescens* (ISA;  $p < 0.05$ ), while *A. lineare* (ISA;  $p < 0.05$ ) was significantly present on lithic substrates.

### 3.3. Multivariate analyses

Non-metric multidimensional scaling (NMS) of diatom assemblages (Fig. 2) well separated communities sampled in the different habitat types: river, springs and peat-bogs (Final stress = 8.09; i.e. good ordination with no real risk of drawing false interference).

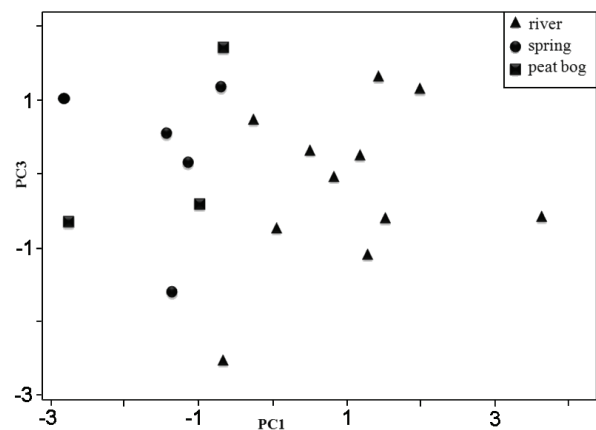


**Fig. 2.** Non-metric multidimensional scaling (NMS) performed on diatom communities.

Multiresponse Permutation Procedure (MRPP) confirmed the NMS findings, showing homogeneity within the three groups ( $A \sim 0.1$ ;  $p < 0.001$ ) and good heterogeneity among them. Moreover, MRPP performed on epi-

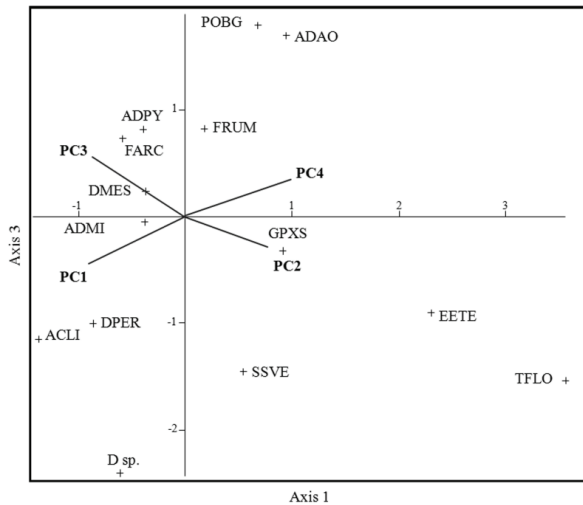
phytic and epilithic assemblages did not show significant separation between communities ( $A = 0.004$ ;  $p = 0.27$ ). Indeed, there were no significant differences in diatom composition between epilithic and epiphytic assemblages.

Considering the small range of variation of the environmental parameters evaluated in the sampled area, we created a further environmental matrix summarizing all the parameters through a Principal Component Analysis (see Fig. 3 for PCA plot). Four significant components were obtained: PC1 explained 28.7% of variance and was mainly related to pH (0.55) and water velocity (0.42). PC2 explained 22.58% of variance and was related to conductivity (-0.55) and Ca (-0.54). PC3 explained 12.72% and was related to dissolved oxygen (-0.70). PC4 explained 11.39% and was related to P-PO<sub>4</sub> (0.53).



**Fig. 3.** Graphical representation of the Principal Component Analysis (PCA).

The coordinates (scores) of the plot were used as an environmental matrix to perform a Canonical Correspondence Analysis. The biological matrix was created by merging epiphytic and epilithic samples coming from the same site, excluding taxa detected in less than 50% of the samples and with less than 5% abundance (the final matrix included 14 taxa). The CCA summary statistics showed a high cumulative percentage of explained variance (38.8%), mostly due to the first axis (24.8%). The graphical representation of CCA is presented in figure 4: only axes 1 and 3 were statistically significant (Monte Carlo test, 99 runs). In general, the left of the graph contains rheophilous diatom species typical of circumneutral pH (PC1), while the right side contains acidophilous taxa, e.g. *Eunotia exigua* var. *tentella* and *T. flocculosa*. We found a good separation between *Diatoma mesodon* and *Diatoma* sp. in terms of ecological preferences: *D. mesodon*, lying in the centre of the graph, shows a more generalist behaviour than *Diatoma* sp. The latter was strongly correlated to PC1, preferring slightly basic pH and lotic conditions, but negatively correlated to PC4, i.e. mainly phosphate concentrations, thus appearing as a xenosaprobous taxon.



**Fig. 4.** Graphical representation of the Canonical Correspondence Analysis (CCA). See Annex 1 for abbreviations.

#### 4. DISCUSSION

The water quality of the investigated sites was very high, with chemical and environmental features typical of mountain streams in pristine areas. The highest value of BOD<sub>5</sub> was recorded in a peat bog, probably due to the grazing impact and static flow. Sampling sites were characterized by permanence of water, so that hydrological stability was also comparable among sampling sites and substrates. Low Ca concentrations reflected the siliceous nature of the substrate. As pointed out in other studies, when the geology and human pressure are equal within a sampling area, current velocity is the most important and selective parameter influencing diatom community composition (Sabater & Roca 1990; Cantonati & Spitale 2009). In our study, the pH and current velocity were the only parameters varying among the three habitat types and they were the driving force influencing diatom community composition. Obviously, pH and water velocity were significantly lower in peat bogs, with the flow often below the detection limit.

The high species richness of the Valasco Valley (174 taxa recorded at 19 sampling sites) is comparable with data obtained from other high mountain protected areas (131 taxa in 21 samples, Cantonati & Spitale 2009; 98 taxa in 6 samples; Battegazzore *et al.* 2004; 188 taxa at 39 sites Fránková *et al.* 2009) but it is higher than the value obtained from routine monitoring in non-protected areas (88 taxa in 51 samples, Mobili *et al.* 2008). The most frequent and abundant species were *A. minutissimum* s.l. and *D. mesodon*, characteristic of all three habitat types (especially springs), in agreement with studies carried out in comparable conditions and geology (Cantonati 1998b; Cantonati *et al.* 2007). In accordance with Cantonati *et al.* (2007), we found high percentages of *Brachysira brebissonii*, *Eunotia minor* and *T. flocculosa*, especially in peat bogs, while *A.*

*pyrenaicum* was less abundant. Species recently described as typical of springs on carbonate substrates, such as *Achnanthydium dolomiticum* (Cantonati & Lange-Bertalot 2006), *Cymbella tridentina* (Cantonati *et al.* 2010), *Geissleria gereckeii* (Cantonati & Lange-Bertalot 2009) and *Encyonema sublangebertalotii* (Cantonati & Lange-Bertalot 2010), were not found at all, confirming their strong affinity for limestone.

The characteristic assemblages found in our study (Tab. 2) were mainly composed of xeno- and oligosaprobious taxa and can be considered reference communities. The comparison between these potential communities and those found in monitoring sites could be a good tool for the evaluation of human pressure in high mountain streams. Indeed, by comparing diatom community compositions in high mountain streams not included in a protected area and heavily modified from the physical point of view, we noticed that the communities in the Valasco Valley generally lacked  $\beta$ -mesosaprobious taxa such as *Cocconeis placentula*, *Nitzschia dissipata*, *Nitzschia fonticola* or  $\alpha$ -mesosaprobious species such as *Diatoma ehrenbergii*.

Indicator species of river communities were *F. arcus*, a rheophilous taxon, and *D. hyemalis*, typical of siliceous substrates (Cantonati 1998b). *A. lineare* preferred lithic substrates to plant ones (see ISA) and was generally correlated with higher water velocity and negatively correlated with orthophosphates (see CCA). Another interesting result of the CCA was the definition of the autoecological preferences of *Diatoma* sp., already recorded in other areas of the Maritime Alps Natural Park (unpublished data) but never detected in other streams of NW Italy (including Piedmont, Aosta Valley and Liguria). The CCA results indicated that *Diatoma* sp. is markedly stenoeccious: indeed, it seemed to be a xenosaprobious taxon, adapted to living on siliceous substrates and low electrolyte contents, preferring good dissolved oxygen levels and plants as substrates. Further taxonomic investigations, both with traditional methods and recently developed techniques (such as geometric morphometric analysis; Falasco *et al.* 2009b; Novais *et al.* 2009), are necessary to clarify the nature of this taxon.

The biodiversity indices were lower in the river than in springs and peat bogs: the mean Shannon index in springs was 2.34, in accordance with Cantonati *et al.* (2007). Biodiversity was generally higher in the Valasco Valley than in other high-altitude streams not included in a protected area and affected by human pressure (Bona *et al.* 2008).

CCA and ISA revealed characteristic taxa of peat bogs: *A. pfaffiana*, *E. exigua* var. *tenella* and *T. flocculosa*, i.e. colonial species generally inhibited in lotic headwaters, with preferences for low pH (van Dam *et al.* 1994). Moreover, *N. virescens* was abundant in the Valasco Valley, mainly preferring springs and peat bogs and plants as substrate.

The characteristic physical and chemical stability of springs and the presence of mosses and aquatic plants, thereby increasing the heterogeneity of microhabitats, allowed colonization by the highest number of taxa in all our samples and confirmed the role of springs as hot spots for biodiversity (Cantonati *et al.* 2006; Fránková *et al.* 2009). We observed no significant differences in species richness or biodiversity values between epiphytic and epilithic assemblages, confirming the observation of Bertrand *et al.* (2004). However, it is noteworthy that the number of species detected in epiphytic and epilithic samples was significantly higher than in samples from stones alone. About 17% of the recorded species were exclusively found on plant substrates. As pointed out by Cantonati *et al.* (2007), sampling of the dominant substrates (stones, mosses, surface sediment) would be an important improvement for sampling guidelines in high-altitude streams, which presently include only sampling on boulders. The sampling of plants in lithic substrates would add important data on the presence and distribution of taxa.

The highest percentage of diatom taxa included as endangered or decreasing in the German Red List (Lange-Bertalot & Steindorf 1996) was found in peat bogs and springs. In general, endangered species showed low abundances and frequencies, the only exceptions being *Amphora inariensis* and *Stauroforma exiguiformis* (found at 16% of the sites), *E. steineckii* and *Neidium alpinum* (19% of the samples), *E. botuliformis* (25% of the sites). *S. exiguiformis* reached its abundance peak in one spring (6%). However, comparison with the Red List could present some limitations. The lack of historical data on diatom taxa distribution and abundance in NW Italy prevented direct comparison with the Italian flora, leading to uncertainty in species classification, especially for the category "in decrease". In addition, some species listed as endangered in the Red List (for example, *A. inariensis* and *N. angusta*) seem to be frequent and well distributed in our region, as confirmed by previous studies (Batteggazzore *et al.* 2004; Falasco *et al.* 2008). Moreover, the Red List has not been updated since 1996 and the percentage of taxa whose conservation status is unknown is still high. This demonstrates the importance of local and specific studies in remote areas in order to update the Red List and validate it with data from the Alpine region.

## 5. CONCLUSIONS

Despite the small sampling area, about 34% of the species were recorded only at one sampling station, underlining the importance of the preservation of every aquatic habitat of the valley, especially peat bogs. This finding of high species diversity in small habitats confirms the results of other studies, e.g. Bertrand *et al.* (2004) or Cantonati & Spitale (2009). The only impact by humans affecting the Valasco Valley is pasture, and there is a substantial presence of grazing domestic herds

during the summer, but the chemistry of the aquatic ecosystems has apparently not been altered. Herd trampling can represent a mechanical disturbance, compressing and eroding peat bogs, whose recovery is particularly slow (Bracco *et al.* 2004). The protection of the most sensitive habitats, such as peat bogs, with temporary fences is recommended. Constructions of passageways for hikers could protect the sensitive habitats and would make visitors more aware about habitat conservation and lessen the damage from herds.

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## A P P E N D I X

**Appendix** - Complete checklist of all the taxa detected in the samples. R.L. column refers to conservation status defined in Lange-Bertalot & Steindorf (1996): 1 = threatened with extinction. 2 = severely endangered. 3 = endangered. D = data scarce. G = probably endangered. R = rare. V = decreasing. \* = at present not considered threatened. \*\* = surely not threatened. pH column represents taxa sensitivity to acidification (van Dam *et al.* 1994): 1= acidobiontic (optimal occurrence at pH <5.5); 2= acidophilous (mainly occurring at pH <7); 3= circumneutral (mainly occurring at pH values about 7); 4= alkaliphilous (mainly occurring at pH >7); 5= alkalibiontic (exclusively occurring at pH >7); 6= indifferent. Moist. column represents taxa moisture preferences (van Dam *et al.* 1994): 1= never or very rarely occurring outside water bodies; 2= mainly occurring in water bodies, sometimes on wet places; 3= mainly occurring in water bodies, also rather regularly on wet and moist places; 4= mainly occurring on wet and moist or temporarily dry places; 5= nearly exclusively occurring outside water bodies. The last columns represent the presence of each single taxon in the sampled habitats and assemblages.

Taxon	Code	R.L.	pH	Moist	R		S		P	
					EL	EP	EL	EP	EL	EP
<i>Achnanthydium daonense</i> (Lange-Bert) Lange-Bert, Monnier et Ector	ADAO	G	3	1	+	+	+	+	+	+
<i>Achnanthydium lineare</i> W. Smith	ACLI	3	3		+	+	+	+	+	+
<i>Achnanthydium minutissimum</i> (Kützing) Czarnecki	ADMI	**	3	3	+	+	+	+	+	+
<i>Achnanthydium minutissimum</i> (Kützing) Czarnecki fo. teratogene	ADMT	1	3	3	+					
<i>Achnanthydium minutissimum</i> var. <i>affinis</i> (Grunow) Bukht.	ADMF	*	4	1	+		+			+
<i>Achnanthydium pyrenaicum</i> (Hustedt) Kobayasi	ADPY	**	4		+	+	+			+
<i>Achnanthydium pyrenaicum</i> (Hustedt) Kobayasi abnormal form	ADPT	1	4		+	+				
<i>Adlafia minuscula</i> var. <i>muralis</i> (Grunow) Lange-Bertalot	ADMM	**	4	3	+	+				
<i>Amphora inariensis</i> Krammer	AINA	3			+	+				+
<i>Amphora ovalis</i> (Kützing) Kützing	AOVA	**	4	1			+			
<i>Amphora pediculus</i> (Kützing) Grunow	APED	**	4	3	+	+				
<i>Aulacoseira alpigena</i> (Grunow) Krammer	AUAL	G	2	1	+		+	+	+	+
<i>Aulacoseira distans</i> (Ehrenberg) Simonsen	AUDI	G	2	1		+				
<i>Aulacoseira pfaffiana</i> (Reinsch) Krammer	AUPF	G	2	2	+		+	+	+	+
<i>Brachysira brebissonii</i> Ross	BBRE	*	2	3		+	+	+	+	+
<i>Brachysira neoexilis</i> Lange-Bertalot	BNEO	*	2	2	+	+	+			
<i>Caloneis tenuis</i> (Gregory) Krammer	CATE	G	3	4	+		+	+		
<i>Cavinula pseudoscutiformis</i> (Hustedt) Mann & Stickle	CPSE	3	4	3	+				+	+
<i>Chamaepinnularia soehrensii</i> var. <i>hassiaci</i> (Krasske) Lange-Bertalot	CHSH	3	2	4				+		+
<i>Cocconeis euglypta</i> Ehrenberg	CPLE	**	4	2				+		
<i>Cocconeis neodiminuta</i> Krammer	CNDI	R				+				
<i>Cocconeis pediculus</i> Ehrenberg	CPED	**	4	1				+		
<i>Cocconeis placentula</i> Ehrenberg	CPLA	4	2	2	+	+				
<i>Cocconeis pseudolineata</i> (Geitler) Lange-Bertalot	CPPL	D	4		+	+				+
<i>Cyclotella atomus</i> Hustedt	CATO	**	4	1						+
<i>Cyclotella ocellata</i> Pantocsek	COCE	*	4	1					+	
<i>Cymbella excisa</i> Kützing	CAEX		4	2	+			+		
<i>Cymbella excisiformis</i> Krammer	CEXF		4		+					
<i>Cymbella hantzschiana</i> Krammer	CHAN				+					
<i>Cymbella helvetica</i> Kützing	CHEL	V	4	2	+					
<i>Cymbella laevis</i> Naegeli in Kützing	CLAE	G	3		+					
<i>Cymbella parva</i> (W. Smith) Kirchner in Cohn	CPAR				+					
<i>Cymbopleura amphicephala</i> Krammer	CBAM	V	3	3				+		
<i>Cymbopleura naviculiformis</i> (Auerswald) Krammer	CBNA	*	3	2		+	+			+
<i>Delicata delicatula</i> (Kützing) Krammer	DDEL	G	4	3				+		
<i>Denticula kuetzingii</i> Grunow	DKUE	*	4	3	+					
<i>Denticula tenuis</i> Kützing	DTEN	*	4	3	+	+	+			
<i>Diadsmis biceps</i> Arnott ex Grunow	DBIC		4	4			+			
<i>Diadsmis perpusilla</i> (Grunow) D.G. Mann	DPER	**	3	5	+	+	+	+		
<i>Diatoma ehrenbergii</i> Kützing	DEHR	**	5	1			+			+
<i>Diatoma hyemalis</i> (Roth) Heiberg	DHIE	*	4	2	+	+		+		
<i>Diatoma mesodon</i> Kützing	DMES	*	3	2	+	+	+	+	+	+
<i>Diatoma moniliformis</i> Kützing	DMON	**	5		+		+			
<i>Diatoma</i> sp.	D sp.				+	+	+	+	+	+

(continued)

## Appendix. Continuation

Taxon	Code	R.L.	pH	Moist	R		S		P	
					EL	EP	EL	EP	EL	EP
<i>Diatoma</i> sp. <i>teratologica</i>	D sp.t									
<i>Encyonema caespitosum</i> Kützing	ECAE	**				+	+			
<i>Encyonema lange-bertalotii</i> Krammer morphotype 1	ENLB		3					+		
<i>Encyonema lunatum</i> (W. Smith) Van Heurck	ENLU						+			+
<i>Encyonema minutum</i> (Hilse) Mann	ENMI	*	3			+	+	+	+	
<i>Encyonema minutum</i> (Hilse in Rabh.) Mann fo. <i>teratogene</i>	ENMT	1				+				
<i>Encyonema neogracile</i> Krammer	ENNG	3	2	3		+	+	+	+	
<i>Encyonema silesiacum</i> (Bleisch) Mann	ESLE	*	3	1		+	+	+	+	+
<i>Encyonema ventricosum</i> (Kützing) Grunow	ENVE	*	3			+	+			
<i>Encyonopsis cesatii</i> (Rabenhorst) Krammer	ECES	*	3	3		+		+		
<i>Encyonopsis falaisensis</i> (Grunow) Krammer	ECFA	G	0	3		+	+	+	+	
<i>Encyonopsis subminuta</i> Krammer et Reichardt	ESUM		3			+				
<i>Encyonopsis minuta</i> Krammer et Reichardt	ECPM		4			+	+	+	+	+
<i>Eolimna minima</i> (Grunow) Lange-Bertalot	EOMI	**	4	3		+	+	+	+	+
<i>Eolimna subminuscula</i> (Manguin) Moser Lange-Bertalot et Metzeltin	ESBM	**	4	3		+				
<i>Eucocconeis laevis</i> (Østrup) Lange-Bertalot	EULA	*	3	3		+	+			
<i>Eunotia borealpina</i> Lange-Bertalot et Nörpel-Schempp	EBOA					+	+	+	+	+
<i>Eunotia botuliformis</i> Wild, Nörpel-Schempp et Lange-Bertalot	EBOT	2	2			+	+	+	+	+
<i>Eunotia diodon</i> Ehrenberg	EDIO	*	2	4						+
<i>Eunotia exigua</i> var. <i>tenella</i> (Grunow) Nörpel et Alles	EETE	V	2	3		+	+	+	+	+
<i>Eunotia glacialisfalsa</i> Lange-Bertalot	EGFA							+		+
<i>Eunotia implicata</i> Nörpel-Schempp, Lange-Bertalot et Alles	EIMP	G	2	3		+	+	+		
<i>Eunotia incisa</i> Gregory	EINC	*	2	2				+		+
<i>Eunotia inflata</i> (Grunow) Nörpel-Schempp et Lange-Bertalot	EINF	2	2					+		+
<i>Eunotia meisterioides</i> Lange-Bertalot										+
<i>Eunotia minor</i> (Kützing) Grunow	EMIN	*	2	4		+	+	+	+	+
<i>Eunotia neocompacta</i> Mayama	ENEC									+
<i>Eunotia paludosa</i> Grunow in Van Heurck	EUPA	V	1	4				+		
<i>Eunotia pseudogroenlandica</i> Lange-Bertalot & Tagliaventi								+		
<i>Eunotia soleirolii</i> (Kützing) Rabenhorst	ESOL		3	3				+		+
<i>Eunotia steineckii</i> Petersen	ESTK	2	2			+			+	+
<i>Eunotia subarcuatoidea</i> Alles, Nörpel et Lange-Bertalot	ESUB	**	1	3				+		+
<i>Eunotia tetraodon</i> Ehrenberg	ETET	2	2	3						+
<i>Eunotia valida</i> Hustedt	EVAL	G	2	3			+			
<i>Fragilaria</i> cf. <i>alpestris</i> Krasske ex Hustedt	FALP	V		4		+	+	+		+
<i>Fragilaria arcus</i> (Ehrenberg) Cleve	FARC	**	4	3		+	+	+		+
<i>Fragilaria arcus</i> (Ehrenberg) Cleve fo. <i>teratogene</i>	FART	1	4	3		+	+	+		+
<i>Fragilaria delicatissima</i> (W. Smith) Lange-Bertalot	FDEL	V	3	1			+			
<i>Fragilaria gracilis</i> (Østrup) Hustedt	FGRA	*	3			+		+		+
<i>Fragilaria rumpens</i> (Kützing) Carlson	FRUM	*	3			+	+	+	+	+
<i>Fragilaria rumpens</i> (Kützing) Carlson fo. <i>teratogene</i>	FRUT	1	3			+	+	+		+
<i>Fragilaria vaucheriae</i> (Kützing) Peterson	FVAU	**	4	3			+	+		
<i>Fragilaria vaucheriae</i> (Kützing) Peterson fo. <i>teratogene</i>	FCVT	1	4	3				+		
<i>Frustulia crassinervia</i> (Brebisson) Lange-Bertalot et Krammer	FCRS	V	1	3		+	+	+	+	+
<i>Gomphonema</i> aff. <i>affine</i> Kützing	GAFF		4	2		+	+	+	+	+
<i>Gomphonema cymbelliclinum</i> Reichardt et Lange-Bertalot	GCBC		4					+		
<i>Gomphonema gracile</i> Ehrenberg	GGRA	D	3	3				+	+	+
<i>Gomphonema micropus</i> Fricke	GMIC	*	3			+				
<i>Gomphonema olivaceoides</i> Hustedt	GOLD	*	3	3		+		+		
<i>Gomphonema olivaceum</i> var. <i>minutissimum</i> Hustedt	GOMI	*	3	3		+	+			
<i>Gomphonema parvulum</i> (Kützing) Kützing	GPAR		3	3		+				
<i>Gomphonema parvulum</i> var. <i>exilissimum</i> Grunow	GPXS	V	3	3		+	+	+	+	+
<i>Gomphonema pseudotenellum</i> Lange-Bertalot	GPTE	3				+				
<i>Gomphonema pumilum</i> var. <i>elegans</i> Reichardt et Lange-Bertalot	GPEL					+	+	+		+
<i>Gomphonema tergestinum</i> (Grunow) Fricke	GTER	G	4	3		+		+		+
<i>Gomphonema utae</i> Lange-Bertalot et Reichardt	GUTA	D						+		+
<i>Kobayasiella parasubrilissima</i> (Kobayasi & Nagumo) Lange-Bertalot	KOPA		1	3						+
<i>Meridion circulare</i> (Greville) Agardh	MCIR	**	4	1				+	+	+
<i>Meridion circulare</i> var. <i>constrictum</i> (Ralfs) Van Heurck	MCCO	**	4	2			+			
<i>Navicula angusta</i> Grunow	NAAN	3	2	2		+	+			+
<i>Navicula cryptotenella</i> Lange-Bertalot	NCTE		4	2		+				
<i>Navicula cryptotenelloides</i> Lange-Bertalot	NCTO	*	4				+			
<i>Navicula exilis</i> Kützing	NEXI	G				+	+			+
<i>Navicula gregaria</i> Donkin	NGRE	**	4	3		+				
<i>Naviculadicta detenta</i> Hustedt	NDET	1				+				+
<i>Naviculadicta difficillima</i> Hustedt	NDIF	G	2	3						+
<i>Naviculadicta parabryophila</i> Lange-Bertalot	NDPA							+		+
<i>Naviculadicta schmassmannii</i> (Hustedt) Werum et Lange-Bertalot	NSMM	2	3			+				
<i>Naviculadicta suchlandtii</i> Hustedt	NSUC	V	3					+	+	+
<i>Neidium alpinum</i> Hustedt	NALP	3	2	3		+	+	+	+	+
<i>Neidium bisulcatum</i> (Lagerstedt) Cleve	NBIS	3	3	3						+
<i>Neidium longiceps</i> (Gregory) Ross	NLGI	G	2	3				+	+	+
<i>Neofragilaria virescens</i> (Ralfs) Williams & Round	NFVI		3			+	+	+	+	+

(continued)

## Appendix. Continuation

Taxon	Code	R.L.	pH	Moist	R		S		P	
					EL	EP	EL	EP	EL	EP
<i>Nitzschia</i> cf. <i>acidoclinata</i> Lange-Bertalot	NACD	*	3	3	+	+	+	+	+	+
<i>Nitzschia</i> cf. <i>bryophila</i> Hustedt	NIBR	D		4			+	+		
<i>Nitzschia</i> <i>dissipata</i> (Kützing) Grunow	NDIS	**	4	3	+	+				
<i>Nitzschia</i> cf. <i>elegantula</i> Grunow	NELE	*						+		
<i>Nitzschia</i> cf. <i>fonticola</i> (Grunow) Grunow	NFON	**	4	1	+		+		+	+
<i>Nitzschia</i> <i>gracilis</i> Hantzsch	NIGR	*	3	1			+	+		
<i>Nitzschia</i> aff. <i>hantzschiana</i> Rabenhorst	NHAN	*	3	4	+	+	+	+	+	+
<i>Nitzschia</i> <i>inconspicua</i> Grunow	NINC	**	4	3	+				+	
<i>Nitzschia</i> <i>obtusa</i> var. <i>brevissima</i> (Grunow) Van Heurck	NOBM							+		
<i>Nitzschia</i> <i>perminuta</i> (Grunow) Peragallo	NIPM	*	4	3	+		+	+		
<i>Nitzschia</i> <i>sinuata</i> var. <i>tabellaria</i> Grunow	NSIT	V	3	3			+			
<i>Nitzschia</i> sp.1						+				
<i>Nupela lapidosa</i> (Krasske) Lange-Bertalot	NULA	V	2	4	+	+	+			+
<i>Orthoseira dentroteres</i> (Ehrenberg) Round, Crawford & Mann	ODEN	V			+	+				
<i>Pinnularia borealis</i> Ehrenberg	PBOR	**	3	4	+	+	+	+	+	+
<i>Pinnularia borealis</i> var. <i>tenuistriata</i> Krammer	PBTE		3	4						+
<i>Pinnularia borealis</i> var. <i>sublinearis</i> Krammer	PBSL		3	4						
<i>Pinnularia frauenbergiana</i> Reichardt	PFRA	R				+				
<i>Pinnularia marchica</i> Ilka Schönfelder var. <i>marchica</i>	PMCH									+
<i>Pinnularia microstauron</i> (Ehrenberg) Cleve	PMIC	V	3	3	+				+	+
<i>Pinnularia microstauron</i> var. <i>rostrata</i> Krammer	PMRO								+	+
<i>Pinnularia peracuminata</i> Krammer	PPEA						+			
<i>Pinnularia perrirorata</i> Krammer	PPRI		2		+		+	+	+	+
<i>Pinnularia sinistra</i> Krammer	PSIN	*	2			+	+	+	+	+
<i>Pinnularia</i> sp.1							+			
<i>Pinnularia stomatophora</i> (Grunow) Cleve	PSTO	G	2	4	+	+				
<i>Pinnularia subcapitata</i> Gregory	PSCA	*	1	3	+					
<i>Pinnularia subcapitata</i> var. <i>elongata</i> Krammer	PSEL	*	2	3			+			
<i>Pinnularia subcapitata</i> var. <i>subrostrata</i> Krammer	PSSR	*	3	3				+		
<i>Pinnularia subrupesstris</i> Krammer	PSRU	G				+				
<i>Planothidium delicatum</i> (Kützing) Round & Bukhtiyarova	PTDE	*	5	3		+				
<i>Planothidium frequentissimum</i> (Lange-Bertalot) Lange-Bertalot	PTFR	**				+				
<i>Planothidium haynaldii</i> (Schaarschmidt) Lange-Bertalot	PHAY	D	4	3	+					
<i>Planothidium lanceolatum</i> (Brébisson) Lange-Bertalot	PTLA	**	4	3	+		+	+		
<i>Psammothidium bioretii</i> (Germain) Bukhtiyarova et Round	PBIO	V	3	4	+					
<i>Psammothidium levanderi</i> (Hustedt) Bukhtiyarova	PLVD	3	3	3		+				
<i>Psammothidium oblongellum</i> (Oestrup) Van de Vijver	POBG	V	3	3	+	+	+	+	+	+
<i>Psammothidium oblongellum</i> (Oestrup) Van de Vijver, teratogena	AOTG	1	3	3					+	
<i>Psammothidium rossii</i> (Hustedt) Bukhtiyarova et Round	PROS	2	3			+				
<i>Pseudostaurosira robusta</i> Williams & Round	PRBS					+				
<i>Reimeria sinuata</i> (Gregory) Kociolek et Stoermer	RSIN	**	3	3	+	+	+			
<i>Reimeria uniseriata</i> Sala Guerrero & Ferrario	RUNI						+			
<i>Rossithidium nodosum</i> (Cleve) Aboal	RNOD	1	3			+				
<i>Rossithidium petersennii</i> (Hustedt) Round & Bukhtiyarova	RPET	3	3		+	+				
<i>Stauriforma exiguiformis</i> (Lange-Bertalot) Flower, V.J. Jones et Round	SEXG	3	3	2	+	+	+	+	+	+
<i>Stauroneis</i> cf. <i>acidoclinata</i> Lange-Bertalot & Werum	SADC				+	+			+	
<i>Stauroneis agrestis</i> Petersen	STAG	R		4	+					
<i>Stauroneis thermicola</i> (Petersen) Lund	STHE	*	3	4			+			
<i>Staurosira construens</i> var. <i>binodis</i> (Ehr.) Hamilton	SCBI	*	4	2		+				
<i>Staurosira pinnata</i> Ehrenberg	SPIN	**	4	3	+		+			
<i>Staurosira venter</i> (Ehrenberg) Kobayasi	SSVE	**	4	1	+	+	+	+	+	+
<i>Surirella brebissonii</i> var. <i>kuetzingii</i> Krammer et Lange-Bertalot	SBKU	**	4	3						
<i>Tabellaria flocculosa</i> (Roth) Kützing	TFLO	**	2	3	+	+	+	+	+	+
<i>Tabellaria flocculosa</i> (Roth) Kützing fo. teratogene	TFLT	1	2	3						
<i>Ulnaria acus</i> (Kützing) Aboal	UACU						+			
<i>Ulnaria ulna</i> (Nitzsch) Compère	UULN	*	4	2	+	+	+			

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